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**UTILITY  
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(Only for new nonprovisional applications under 37 CFR 1.53(b))

Attorney Docket No. 5763-029 Total Pages 77

First Named Inventor or Application Identifier

Michal Eisenbach-Schwartz

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See MPEP chapter 600 concerning utility patent application contents.

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*(preferred arrangement set forth below)*
  - Descriptive title of the Invention
  - Cross Reference to Related Applications
  - Statement Regarding Fed sponsored R&D
  - Reference to Microfiche Appendix
  - Background of the Invention
  - Brief Summary of the Invention
  - Brief Description of the Drawings *(if filed)*
  - Detailed Description of the Invention (including drawings, *if filed*)
  - Claim(s)
  - Abstract of the Disclosure
3.  Drawing(s) (35 USC 113) (17 Figures/25 Sheets) [Total Sheets 25]
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    - i.  **DELETION OF INVENTOR(S)**  
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ATTORNEY DOCKET NO. 5763-029Date December 22, 1998

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Sir:

The following utility patent application is enclosed for filing:

Applicant(s): Michal Eisenbach-Schwartz, Irun R. Cohen, Gila  
 Moalem, Pierre Beserman, and Alon Monsonego      Executed on: Inventors unavailable at time of  
 execution

Title of Invention: ACTIVATED T-CELLS, NERVOUS SYSTEM-SPECIFIC ANTIGENS AND THEIR USES

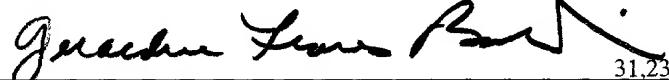
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Total Claims	30	-20	10	\$18.00 each	180.00
Independent	2	-3	0	\$78.00 each	0.00
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Multiple Dependency Fee If Applicable (\$260.00)					260.00
Total					1,200.00
50% Reduction for Independent Inventor, Nonprofit Organization or Small Business Concern (a verified statement as to the applicant's status is attached)					-
Total Filing Fee					\$ 1,200.00

Priority of application no. IL 124550 filed on May 19, 1998 in Israel is claimed under 35 U.S.C. § 119.  
 The certified copy of the priority application has been filed in application no. filed.  
 Amend the specification by inserting before the first line the following sentence; This is a continuation-in-part of application no. filed.

Please charge the required fee to Pennie & Edmonds LLP Deposit Account No. 16-1150. A copy of this sheet is enclosed.

Respectfully submitted,

  
 Geraldine F. Baldwin  
 (Reg. No.)  
 31,232

PENNIE & EDMONDS LLP

Enclosure

This form is not for use with continuation, divisional, re-issue, design or plant patent applications.

**ACTIVATED T-CELLS, NERVOUS SYSTEM-SPECIFIC  
ANTIGENS AND THEIR USES**

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**ACTIVATED T-CELLS, NERVOUS SYSTEM-SPECIFIC  
ANTIGENS AND THEIR USES**

The present application is a continuation-in-part of  
5 PCT/US98/14715, filed July 21, 1998. The present application  
claims priority benefit under 35 U.S.C. § 119 of copending  
Israeli patent application IL 124550, filed May 19, 1998, the  
disclosure of which is incorporated herein by reference in its  
entirety and priority benefit under 35 U.S.C. § 120 of  
10 PCT/US98/14715, filed July 21, 1998.

**1. FIELD OF THE INVENTION**

The present invention relates to compositions and methods  
for the promotion of nerve regeneration or prevention or  
15 inhibition of axon degeneration to ameliorate the effects of  
injury or disease of the nervous system (NS). In certain  
embodiments, activated antiself T-cells, a NS-specific antigen  
or peptide derived therefrom or a nucleotide sequence encoding  
a NS-specific antigen or peptide derived therefrom are/is used  
20 to promote nerve regeneration or to prevent or inhibit axonal  
degeneration caused by injury or disease of nerves within the  
CNS or PNS of a human subject. The compositions of the present  
invention may be administered alone or may be optionally  
administered in any desired combination.

**2. BACKGROUND OF THE INVENTION**

The nervous system comprises the central and the  
peripheral nervous system (PNS). The central nervous system  
(CNS) is composed of the brain and spinal cord; the PNS  
30 consists of all the other neural elements, namely the nerves  
and ganglia outside the brain and spinal cord.

Damage to the NS may result from a traumatic injury, such  
as penetrating trauma or blunt trauma, or a disease or

disorder, including but not limited to Alzheimer's disease, Parkinson's disease, multiple sclerosis, Huntington's disease, amyotrophic lateral sclerosis (ALS), Diabetic neuropathy, senile dementia, and ischemia.

5 Maintenance of CNS integrity is a complex 'balancing act' in which compromises are struck with the immune system. In most tissues, the immune system plays an essential part in protection, repair and healing. In the CNS, because of its unique immune privilege, immunological reactions are relatively  
10 limited (Streilein, J.W., 1993, *Curr. Opin. Immunol.* 5:428-432; Streilein, J.W., 1993, *Science*, 270:1158-1159). A growing body of evidence indicates that the failure of the mammalian CNS to achieve functional recovery after injury is a reflection of an ineffective 'dialog' between the damaged tissue and the immune  
15 system. For example, the restricted communication between the CNS and blood-borne macrophages affects the capacity of axotomized axons to regrow; transplantation of activated macrophages can promote CNS regrowth (Lazarov Spiegler, O., et al., 1996, *FASEB J.*, 10:1296-1302; Rapalino, O. et al., 1998,  
20 *Nature Med.* 4:814-821).

Activated T cells have been shown to enter the CNS parenchyma, irrespective of their antigen specificity, but only T cells capable of reacting with a CNS antigen seem to persist there (Hickey, W.F., et al., 1991, *J. Neurosci. Res.* 28:254-  
25 260). T cells reactive to antigens of CNS white matter, such as myelin basic protein (MBP), can induce the paralytic disease experimental autoimmune encephalomyelitis (EAE) within several days of their inoculation into naive recipient rats (Ben Nun, A., et al., 1981, *Eur. J. Immunol.* 11:195-199). Anti-MBP T  
30 cells may also be involved in the human disease multiple sclerosis (Ota, K., et al., 1990, *Nature* 346:183-187; Martin, R., 1997, *J. Neural Transm. Suppl.* 49:53-67). However, despite

their pathogenic potential, anti-MBP T-cell clones are present in the immune systems of healthy subjects (Burns, J., et al., 1983, *Cell. Immunol.* 81:435-440; Pette, M., et al., 1990, *Proc. Natl. Acad. Sci. USA* 87:7968-7972; Martin, R., et al., 1990, *J. Immunol.* 145:540-548; Schiuesener, H.J., et al., 1985, *J. Immunol.* 135:3128-3133). Activated T cells, which normally patrol the intact CNS, transiently accumulate at sites of CNS white matter lesions (Hirschberg, D.L., et al., 1998, *J. Neuroimmunol.* 89:88-96).

10 A catastrophic consequence of CNS injury is that the primary damage is often compounded by the gradual secondary loss of adjacent neurons that apparently were undamaged, or only marginally damaged, by the initial injury (Faden, A.I., et al., 1992, *Trends Pharmacol. Sci.* 13:29-35; Faden, A.I., 1993, *Crit. Rev. Neurobiol.* 7:175-186; McIntosh, T.K., 1993, *J. Neurotrauma* 10:215-261). The primary lesion causes changes in extracellular ion concentrations, elevation of amounts of free radicals, release of neurotransmitters, depletion of growth factors, and local inflammation. These changes trigger a 20 cascade of destructive events in the adjacent neurons that initially escaped the primary injury (Lynch, D.R., et al., 1994, *Curr. Opin. Neurol.* 7:510-516; Bazan, N.G., et al., 1995, *J. Neurotrauma* 12:791-814; Wu, D., et al., 1994, *J. Neurochem.* 62:37-44). This secondary damage is mediated by activation of 25 voltage-dependent or agonist-gated channels, ion leaks, activation of calcium-dependent enzymes such as proteases, lipases and nucleases, mitochondrial dysfunction and energy depletion, culminating in neuronal cell death (Yoshina, A., et al., 1991, *Brain Res.* 561:106-119; Hovda, D.A., et al., 1991, *Brain Res.* 567:1-10; Zivin, J.A., et al., 1991, *Sci. Am.* 265:56-63; Yoles, E., et al., 1992, *Invest. Ophthalmol. Vis. Sci.* 33:3586-3591). The widespread loss of neurons beyond the 30

loss caused directly by the primary injury has been called 'secondary degeneration'.

Another tragic consequence of CNS injury is that neurons in mammalian CNS do not undergo spontaneous regeneration 5 following an injury. Thus, a CNS injury causes permanent impairment of motor and sensory functions.

Citation or identification of any reference in this section or any other part of this specification shall not be construed as an admission that such reference is available as 10 prior art to the present invention.

### 3. SUMMARY OF THE INVENTION

The present invention is directed to methods and compositions for the promotion of nerve regeneration or 15 prevention or inhibition of axonal degeneration to ameliorate the effects of injury or disease of the nervous system (NS). The present invention is based, in part, on the Applicants' unexpected discovery, that non-recombinant antiself T-cells that recognize an antigen of the NS or a peptide derived 20 therefrom promote nerve regeneration or confer neuroprotection. As used herein, "neuroprotection" refers to the prevention or inhibition of degenerative effects of injury or disease in the NS. Until recently, it was thought that the immune system excluded immune cells from participating in nervous system 25 repair. It was quite surprising to discover that non-recombinant NS-specific antiself activated T-cells can be used to promote nerve regeneration or to protect nervous system tissue from secondary degeneration which may follow damage caused by injury or disease of the CNS or PNS, in particular, 30 a lesion other than a neoplasm or an autoimmune disease affecting the NS.

"Activated T-cell" as used herein includes (i) T-cells

that have been activated by exposure to a cognate antigen or peptide derived therefrom or derivative thereof and (ii) progeny of such activated T-cells. As used herein, a cognate antigen is an antigen that is specifically recognized by the T-5 cell antigen receptor of a T-cell that has been previously exposed to the antigen.

In an embodiment, the present invention provides pharmaceutical compositions comprising a therapeutically effective amount of non-recombinant, NS-specific antiself activated T-cells and methods of use of such compositions for promotion of nerve regeneration or for prevention or inhibition of axonal degeneration in the CNS or PNS in which the amount is effective to ameliorate the effects of an injury or disease of the NS. "NS-specific antiself activated T-cell" as used herein 15 refers to an activated T-cell having specificity for an antigen of the NS or a peptide derived therefrom. Preferably, the NS-specific antiself activated T cells are used to promote nerve regeneration or to prevent or inhibit the effects of disease in which the disease is not an autoimmune disease or a neoplasm.

20 The present invention also provides pharmaceutical compositions comprising a therapeutically effective amount of a NS-specific antigen or peptide derived therefrom or derivative thereof and methods of use of such compositions for promotion of nerve regeneration or for prevention or inhibition 25 of axonal degeneration in the CNS or PNS in which the amount is effective to activate T-cells *in vivo* or *in vitro* wherein the activated T-cells inhibit or ameliorate the effects of an injury or disease of the NS. "NS-specific antigen" as used herein refers to an antigen that specifically activates T-cells 30 such that following activation the activated T-cells accumulate at a site of injury or disease in the NS. Preferably, the NS-specific antigen is used to promote regeneration or to prevent

or inhibit the effects of disease in which the disease is not an autoimmune disease or a neoplasm. In an embodiment, the peptide derived from a NS-specific antigen is a "cryptic epitope" of the antigen. A cryptic epitope activates specific 5 T cells after an animal is immunized with the particular peptide, but not with the whole antigen. In another embodiment, the peptide derived from a NS-specific antigen is an immunogenic epitope of the antigen. "Derivatives" of NS-specific antigens or peptides derived therefrom as used herein 10 refers to analogs or chemical derivatives of such antigens or peptides as described below, see Section 5.2.

The present invention also provides pharmaceutical compositions comprising a therapeutically effective amount of a nucleotide sequence encoding a NS-specific antigen or peptide 15 derived therefrom or derivative thereof and methods of use of such compositions for promotion of nerve regeneration or for prevention or inhibition of axonal degeneration in the CNS or PNS in which the amount is effective to ameliorate the effects of an injury or disease of the NS.

20 In the practice of the invention, therapy for amelioration of effects of injury or disease comprising administration of NS-specific antiself activated T-cells may optionally be in combination with a NS-specific antigen or peptide derived therefrom or derivative thereof or a nucleotide sequence 25 encoding a NS-specific antigen or peptide derived therefrom.

#### 4. BRIEF DESCRIPTION OF THE FIGURES

Fig. 1 shows T-cell presence in injured optic nerve 1 week after injury. Adult Lewis rats were injected with activated T 30 cells of the anti-MBP ( $T_{MBP}$ ), anti-OVA ( $T_{OVA}$ ), or anti-p277 ( $T_{p277}$ ) lines, or with PBS, immediately after unilateral crush injury of the optic nerve. Seven days later, both the injured and

uninjured optic nerves were removed, cryosectioned and analyzed immunohistochemically for the presence of immunolabeled T cells. T cells were counted at the site of injury and at randomly selected areas in the uninjured optic nerves. The 5 histogram shows the mean number of T cells per  $\text{mm}^2 \pm \text{s.e.m.}$ , counted in two to three sections of each nerve. Each group contained three to four rats. The number of T cells was considerably higher in injured nerves of rats injected with anti-MBP, anti-OVA or anti-p277 T cells; statistical analysis 10 (one-way ANOVA) showed significant differences between T cell numbers in injured optic nerves of rats injected with anti-MBP, anti-OVA, or anti-p277 T cells and in injured optic nerves of rats injected with PBS ( $P<0.001$ ); and between injured optic nerves and uninjured optic nerves of rats injected with anti- 15 MBP, anti-OVA, or anti-p277 T cells ( $P<0.001$ ).

Fig. 2 illustrates that T cells specific to MBP, but not to OVA or p277 or hsp60, protect neurons from secondary degeneration. Immediately after optic nerve injury, rats were injected with anti-MBP, anti-OVA or anti-p277 T cells, or with 20 PBS. The neurotracer dye 4-Di-10-Asp was applied to optic nerves distal to the site of the injury, immediately after injury (for assessment of primary damage) or 2 weeks later (for assessment of secondary degeneration). Five days after dye application, the retinas were excised and flat-mounted. 25 Labeled retinal ganglion cells (RGCs) from three to five randomly selected fields in each retina (all located at approximately the same distance from the optic disk) were counted by fluorescence microscopy. RGC survival in each group of injured nerves was expressed as the percentage of the total 30 number of neurons spared after the primary injury (42% of axons remained undamaged after the primary injury). The neuroprotective effect of anti-MBP T cells compared with that

of PBS was significant ( $P<0.001$ , one-way ANOVA). Anti-OVA T cells or anti-p277 T cells did not differ significantly from PBS in their effects on the protection of neurons that had escaped primary injury ( $P>0.05$ , one-way ANOVA). The results 5 are a summary of five experiments. Each group contained five to ten rats.

**Figs. 3(A-C)** present photomicrographs of retrogradely labeled retinas of injured optic nerves of rats. Immediately after unilateral crush injury of their optic nerves, rats were 10 injected with PBS (Fig. 3A) or with activated anti-p277 T cells (Fig. 3B) or activated anti-MBP T cells (Fig. 3C). Two weeks later, the neurotracer dye 4-Di-10-Asp was applied to the optic nerves, distal to the site of injury. After 5 days, the retinas were excised and flat-mounted. Labeled (surviving) 15 RCGs, located at approximately the same distance from the optic disk in each retina, were photographed.

**Figs. 4(A-B)** show that clinical severity of EAE is not influenced by an optic nerve crush injury. For the results presented in Fig. 4A, Lewis rats, either uninjured (dash line) 20 or immediately after optic nerve crush injury (solid line), were injected with activated anti-MBP T cells. EAE was evaluated according to a neurological paralysis scale. [Data points represent  $\pm$  s.e.m.] These results represent a summary of three experiments. Each group contained five to nine rats. 25 Fig. 4B shows that the number of RGCs in the uninjured optic nerve is not influenced by injection of anti-MBP T cells. Two weeks after the injection of anti-MBP T cells or PBS, 4-Di-10-Asp was applied to the optic nerves. After 5 days the retinas were excised and flat-mounted. Labeled RGCs from five fields 30 (located at approximately the same distance from the optic disk) in each retina were counted and the average number per  $\text{mm}^2$  was calculated. There was no difference between the numbers of

labeled RGCs in rats injected with anti-MBP T cells ( $T_{MBP}$ ) and in PBS-injected control rats.

**Fig. 5** shows that T cells specific to p51-70 of MBP protect neurons from secondary degeneration. Immediately after 5 optic nerve injury, rats were injected with anti-MBP T cells, anti-p51-70 T cells, or PBS. The neurotracer dye 4-Di-10-Asp was applied to optic nerves distal to the site of the injury, immediately after injury (for assessment of primary damage) or 2 weeks later (for assessment of secondary degeneration). Five 10 days after dye application, the retinas were excised and flat-mounted. Labeled retinal ganglion cells (RGCs) from three to five randomly selected fields in each retina (all located at approximately the same distance from the optic disk) were counted by fluorescence microscopy. RGC survival in each group 15 of injured nerves was expressed as the percentage of the total number of neurons spared after primary injury. Compared with that of PBS treatment, the neuroprotective effects of anti-MBP and anti-p51-70 T cells were significant ( $P<0.001$ , one-way ANOVA).

20 **Figs. 6(A-B)** show that anti-MBP T cells increase the compound action potential (CAP) amplitudes of injured optic nerves. Immediately after optic nerve injury, rats were injected with either PBS or activated anti-MBP T cells ( $T_{MBP}$ ). Two weeks later, the CAPs of injured (Fig. 6A) and uninjured 25 (Fig. 6B) nerves were recorded. There were no significant differences in mean CAP amplitudes between uninjured nerves obtained from PBS-injected and T cell-injected rats ( $n=8$ ;  $p=0.8$ , Student's t-test). The neuroprotective effect of anti-MBP T cells (relative to PBS) on the injured nerve on day 14 30 after injury was significant ( $n=8$ ;  $p=0.009$ , Student's t-test).

**Fig. 7** illustrates inhibition of secondary degeneration after optic nerve crush injury in adult rats. See text,

Section 8, for experimental details. Rats were injected intradermally through the footpads with a 21-mer peptide based on amino acid residues 35-55 (MOG p35-55) of myelin/oligodendrocyte glycoprotein (chemically synthesized at 5 the Weizmann Institute, Israel) (50 µg/animal) or PBS ten days prior to optic nerve crush injury or MOG p35-55 in the absence of crush injury. MOG p35-55 was administered with Incomplete Freund's Adjuvant. Surviving optic nerve fibers were monitored by retrograde labeling of retinal ganglion cells (RGCs). The 10 number of RGCs in rats injected with PBS or MOG p35-55 was expressed as a percentage of the total number of neurons in rats injected with MOG p35-55 in the absence of crush injury.

Fig. 8 illustrates inhibition in adult rats of secondary degeneration after optic nerve crush injury by MBP. See text, 15 Section 9, for experimental details. MBP (Sigma, Israel) (1 mg in 0.5 ml saline) was administered orally to adult rats by gavage using a blunt needle. MBP was administered 5 times, i.e., every third day beginning 2 weeks prior to optic nerve crush injury. Surviving optic nerve fibers were monitored by 20 retrograde labeling of retinal ganglion cells (RGCs). The number of RGCs in treated rats was expressed as a percentage of the total number of neurons in untreated rats following the injury.

Fig. 9 shows the nucleotide sequence of rat myelin basic 25 protein gene, SEQ ID NO: \_\_\_\_\_, Genbank accession number M25889 (Schaich et al., 1986, *Biol. Chem.* 367, 825-834).

Fig. 10 shows the nucleotide sequence of human myelin basic protein gene, SEQ ID NO: \_\_\_\_\_, Genbank accession number M13577 (Kamholz et al., 1986, *Proc. Natl. Acad. Sci. U.S.A.* 83 (13), 4962-4966).

30 Figs. 11(A-F) show the nucleotide sequences of human myelin proteolipid protein gene exons 1-7, SEQ ID NO: \_\_\_\_\_, Genbank accession numbers M15026-M15032 respectively (Diehl et

al., [published erratum appears in Proc Natl Acad Sci U S A, 1991, 86(6):617-8] *Proc. Natl. Acad. Sci. U.S.A.* 83 (24), 9807-9811 (1986)).

Fig. 12 shows the nucleotide sequence of human myelin oligodendrocyte glycoprotein gene, SEQ ID NO: \_\_\_\_\_, Genbank accession number Z48051 (Roth et al., submitted (17-JAN-1995) Roth, CNRS UPR 8291, CIGH, CHU Purpan, Toulouse, France, 31300; Gonzalez et al., 1996, *Mol. Phylogenet. Evol.* 6, 63-71).

Fig. 13 shows the nucleotide sequence of rat proteolipid protein and variant, SEQ ID NO: \_\_\_\_\_, Genbank accession number M16471 (Nave et al., 1987, *Proc. Natl. Acad. Sci. U.S.A.* 84, 5665-5669).

Fig. 14 shows the nucleotide sequence of rat myelin-associated glycoprotein, SEQ ID NO: \_\_\_\_\_, Genbank accession number M14871 (Arquint et al., 1987, *Proc. Natl. Acad. Sci. U.S.A.* 84, 600-604).

Fig. 15 shows the amino acid sequence of human myelin basic protein, SEQ ID NO: \_\_\_\_\_, Genbank accession number 307160 (Kamholz et al., 1986, *Proc. Natl. Acad. Sci. U.S.A.* 83 (13), 4962-4966).

Fig. 16 shows the amino acid sequence of human proteolipid protein, SEQ ID NO: \_\_\_\_\_, Genbank accession number 387028.

Fig. 17 shows the amino acid sequence of human myelin oligodendrocyte glycoprotein, SEQ ID NO: \_\_\_\_\_, Genbank accession number 793839 (Roth et al., 1995, *Genomics* 28 (2), 241-250; Roth Submitted (17-JAN-1995) Roth CNRS UPR 8291, CIGH, CHU Purpan, Toulouse, France, 31300; Gonzalez et al., 1996, *Mol. Phylogenet. Evol.* 6, 63-71).

##### 5. DETAILED DESCRIPTION OF THE INVENTION

Merely for ease of explanation, the detailed description of the present invention is divided into the following sub-sections: (1) non-recombinant, NS-specific antiself activated

T-cells; (2) NS-specific antigens, peptides derived therefrom and derivatives thereof; (3) nucleotide sequences encoding NS-specific antigens and peptides derived therefrom; (4) therapeutic uses of non-recombinant, NS-specific antiself activated T-cells, NS-specific antigens, peptides derived therefrom and derivatives thereof, and nucleotide sequences encoding NS-specific antigens and peptides derived therefrom; and (5) formulations and modes of administration of non-recombinant, NS-specific antiself activated T-cells, NS-specific antigens, peptides derived therefrom and derivatives thereof, and nucleotide sequences encoding NS-specific antigens and peptides derived therefrom.

#### **5.1 NS-SPECIFIC ANTISELF ACTIVATED T-CELLS**

15 NS-specific antiself activated T-cells (ATCs) can be used for ameliorating or inhibiting the effects of injury or disease of the CNS or PNS that result in NS degeneration or for promoting regeneration in the NS, in particular the CNS.

15 The NS-specific activated T-cells are preferably autologous, most preferably of the CD4 and/or CD8 phenotypes, but they may be also allogeneic T-cells from related donors, e.g. siblings, parents, children, or HLA-matched or partially matched, semi-allogeneic or fully allogeneic donors.

15 The NS-specific antiself activated T-cells are preferably non-attenuated, although attenuated NS-specific activated T-cells may be used. T-cells may be attenuated using methods well known in the art, including but not limited to, by gamma-irradiation, e.g. 1.5-10.0 Rads (Ben-Nun, A., Wekerle, H. and Cohen, I.R., *Nature* 292:60-61 (1981); Ben-Nun, A. and Cohen, I.R., *J. Immunol.* 129:303-308 (1982)); and/or by pressure treatment, for example as described in U.S. Patent No. 4,996,194 (Cohen et al.); and/or by chemical cross-linking with

an agent such as formaldehyde, glutaraldehyde and the like, for example as described in U.S. Patent No. 4,996,194 (Cohen et al.); and/or by cross-linking and photoactivation with light with a photoactivatable psoralen compound, for example as 5 described in U.S. Patent No. 5,114,721 (Cohen et al.); and/or by a cytoskeletal disrupting agent such as cytochalsin and colchicine, for example as described in U.S. Patent No. 4,996,194 (Cohen et al.). In a preferred embodiment the NS-specific antiself activated T-cells are isolated as described 10 below. T-cells can be isolated and purified according to methods known in the art (Mor and Cohen, 1995, *J. Immunol.* 155:3693-3699). For an illustrative example, see Section 6.1.

Circulating T-cells of a subject which recognize myelin basic protein or another NS antigen such as the amyloid 15 precursor protein are isolated and expanded using known procedures. In order to obtain NS-specific antiself activated T-cells, T-cells are isolated and the NS-specific ATCs are then expanded by a known procedure (Burns et al., *Cell Immunol.* 81:435 (1983); Pette et al., *Proc. Natl. Acad. Sci. USA* 87:7968 20 (1990); Mortin et al., *J. Immunol.* 145:540 (1990); Schluesener et al., *J. Immunol.* 135:3128 (1985); Suruhan-Dires Keneli et al., *Euro. J. Immunol.* 23:530 (1993) which are incorporated herein by reference in their entirety.

The isolated T-cells may be activated by exposure of the 25 cells to one or more of a variety of natural or synthetic NS-specific antigens or epitopes, including but not limited to, myelin basic protein (MBP), myelin oligodendrocyte glycoprotein (MOG), proteolipid protein (PLP), myelin-associated glycoprotein (MAG), S-100,  $\beta$ -amyloid, Thy-1, P0, P2 and 30 neurotransmitter receptors. In a preferred embodiment, the isolated T cells are activated by one or more cryptic epitopes, including but limited to the following MBP peptides: p11-30,

p51-70, p91-110, p131-150, and p151-170.

During *ex vivo* activation of the T-cells, the T-cells may be activated by culturing them in medium to which at least one suitable growth promoting factor has been added. Growth 5 promoting factors suitable for this purpose include, without limitation, cytokines, for instance tumor necrosis factor  $\alpha$  (TNF- $\alpha$ ), interleukin 2 (IL-2), and interleukin 4 (IL-4).

In an embodiment, the activated T-cells endogenously produce a substance that ameliorates the effects of injury or 10 disease in the CNS.

In another embodiment, the activated T-cells endogenously produce a substance that stimulates other cells, including, but not limited to, transforming growth factor- $\beta$  (TGF- $\beta$ ), nerve growth factor (NGF), neurotrophic factor 3 (NT-3), neurotrophic 15 factor 4/5 (NT-4/5), brain-derived neurotrophic factor (BDNF); interferon- $\gamma$  (IFN- $\gamma$ ), interleukin-6 (IL-6), wherein the other cells, directly or indirectly, ameliorate the effects of injury or disease.

Following their proliferation *in vitro*, the T-cells are 20 administered to a mammalian subject. In a preferred embodiment, the T-cells are administered to a human subject. T-cell expansion is preferably performed using peptides corresponding to sequences in a non-pathogenic, NS-specific, self protein.

25 A subject can initially be immunized with a NS-specific antigen using a non-pathogenic peptide of the self protein. A T-cell preparation can be prepared from the blood of such immunized subjects, preferably from T-cells selected for their specificity towards the NS-specific antigen. The selected T- 30 cells can then be stimulated to produce a T-cell line specific to the self-antigen (Ben-Nun *et al.*, *J. Immunol.* 129:303 (1982)).

The NS-specific antigen may be a purified antigen or a crude NS preparation, as will be described below.

NS-specific antigen activated T-cells, obtained as described above, can be used immediately or may be preserved 5 for later use, e.g. by cryopreservation as described below. NS-specific antiself activated T-cells may also be obtained using previously cryopreserved T-cells, i.e., after thawing the cells, the T-cells may be incubated with NS-specific antigen, optimally together with thymocytes, to obtain a preparation of 10 NS-specific ATCs.

As will be evident to those skilled in the art, the T-cells can be preserved, e.g. by cryopreservation, either before or after culture.

Cryopreservation agents which can be used include but are 15 not limited to dimethyl sulfoxide (DMSO) (Lovelock and Bishop, 1959, *Nature* 183:1394-1395; Ashwood-Smith, 1961, *Nature* 190:1204-1205), glycerol, polyvinylpyrrolidone (Rinfret, 1960, *Ann. N.Y. Acad. Sci.* 85:576), polyethylene glycol (Sloviter and Ravdin, 1962, *Nature* 196:548), albumin, dextran, sucrose, 20 ethylene glycol, i-erythritol, D-ribitol, D-mannitol (Rowe et al., 1962, *Fed. Proc.* 21:157), D-sorbitol, i-inositol, D-lactose, choline chloride (Bender et al., 1960, *J. Appl. Physiol.* 15:520), amino acids (Phan The Tran and Bender, 1960, *Exp. Cell Res.* 20:651), methanol, acetamide, glycerol 25 monoacetate (Lovelock, 1954, *Biochem. J.* 56:265), inorganic salts (Phan The Tran and Bender, 1960, *Proc. Soc. Exp. Biol. Med.* 104:388; Phan The Tran and Bender, 1961, in *Radiobiology, Proceedings of the Third Australian Conference on Radiobiology*, Ilbery, P.L.T., ed., Butterworth, London, p. 59), and DMSO 30 combined with hydroxyethyl starch and human serum albumin (Zaroulis and Leiderman, 1980, *Cryobiology* 17:311-317).

A controlled cooling rate is critical. Different

cryoprotective agents (Rapatz et al., 1968, *Cryobiology* 5(1):18-25) and different cell types have different optimal cooling rates. See, e.g., Rowe and Rinfret, 1962, *Blood* 20:636; Rowe, 1966, *Cryobiology* 3(1):12-18; Lewis et al., 1967, *Transfusion* 7(1):17-32; and Mazur, 1970, *Science* 168:939-949 for effects of cooling velocity on survival of cells and on their transplantation potential. The heat of fusion phase where water turns to ice should be minimal. The cooling procedure can be carried out by use of, e.g., a programmable freezing device or a methanol bath procedure.

Programmable freezing apparatuses allow determination of optimal cooling rates and facilitate standard reproducible cooling. Programmable controlled-rate freezers such as Cryomed or Planar permit tuning of the freezing regimen to the desired cooling rate curve.

After thorough freezing, cells can be rapidly transferred to a long-term cryogenic storage vessel. In one embodiment, samples can be cryogenically stored in mechanical freezers, such as freezers that maintain a temperature of about -80°C or 20 about -20°C. In a preferred embodiment, samples can be cryogenically stored in liquid nitrogen (-196°C) or its vapor. Such storage is greatly facilitated by the availability of highly efficient liquid nitrogen refrigerators, which resemble large Thermos containers with an extremely low vacuum and 25 internal super insulation, such that heat leakage and nitrogen losses are kept to an absolute minimum.

Considerations and procedures for the manipulation, cryopreservation, and long term storage of T-cells can be found, for example, in the following references, incorporated by reference herein: Gorin, 1986, Clinics in Haematology 30 15(1):19-48; Bone-Marrow Conservation, Culture and Transplantation, Proceedings of a Panel, Moscow, July 22-26,

1968, International Atomic Energy Agency, Vienna, pp. 107-186.

Other methods of cryopreservation of viable cells, or modifications thereof, are available and envisioned for use, e.g., cold metal-mirror techniques. See Livesey and Linner, 5 1987, Nature 327:255; Linner et al., 1986, J. Histochem. Cytochem. 34(9):1123-1135; see also U.S. Patent No. 4,199,022 by Senken et al., U.S. Patent No. 3,753,357 by Schwartz, U.S. Patent No. 4,559,298 by Fahy.

Frozen cells are preferably thawed quickly (e.g., in a 10 water bath maintained at 37-41°C) and chilled immediately upon thawing. It may be desirable to treat the cells in order to prevent cellular clumping upon thawing. To prevent clumping, various procedures can be used, including but not limited to the addition before or after freezing of DNase (Spitzer et al., 15 1980, Cancer 45:3075-3085), low molecular weight dextran and citrate, hydroxyethyl starch (Stiff et al., 1983, Cryobiology 20:17-24), or acid citrate dextrose (Zaroulis and Leiderman, 1980, Cryobiology 17:311-317), etc.

The cryoprotective agent, if toxic in humans, should be 20 removed prior to therapeutic use of the thawed T-cells. One way in which to remove the cryoprotective agent is by dilution to an insignificant concentration.

Once frozen T-cells have been thawed and recovered, they are used to promote axonal regeneration as described herein 25 with respect to non-frozen T-cells.

#### **5.2 NS-SPECIFIC ANTIGENS AND PEPTIDES DERIVED THEREFROM**

Pharmaceutical compositions comprising a NS-specific antigen or peptide derived therefrom or derivative thereof can 30 be used for preventing or inhibiting the effects of injury or disease that result in NS degeneration or for promoting nerve regeneration in the NS, particularly in the CNS. Additionally,

NS-specific antigens or peptides derived therefrom or derivatives thereof may be used for *in vivo* or *in vitro* activation of antiself T-cells. In an embodiment, the NS-specific antigen is an isolated or purified antigen. In an 5 embodiment, methods of promoting nerve regeneration or of preventing or inhibiting the effects of CNS or PNS injury or disease comprise administering NS-specific antigen or a peptide derived therefrom or derivative thereof to a mammal wherein the NS-specific antigen or peptide derived therefrom or derivative 10 thereof activates T-cells *in vivo* to produce a population of T-cells that accumulate at a site of injury or disease of the CNS or PNS.

The NS-specific antigen may be an antigen obtained from NS tissue, preferably from tissue at a site of CNS injury or 15 disease. The NS-specific antigen may be isolated and purified by standard methods including chromatography (e.g., ion exchange, affinity, and sizing column chromatography), centrifugation, differential solubility, or by any other standard technique for the purification of antigens. The 20 functional properties may be evaluated using any suitable assay. In the practice of the invention, natural or synthetic NS-specific antigens or epitopes include, but are not limited to, MBP, MOG, PLP, MAG, S-100,  $\beta$ -amyloid, Thy-1, P0, P2 and a neurotransmitter receptor.

25 Specific illustrative examples of useful NS-specific antigens include but are not limited to, human MBP, depicted in Fig. 15 (SEQ ID NO: \_\_\_\_\_); human proteolipid protein, depicted in Fig. 16 (SEQ ID NO: \_\_\_\_\_); and human oligodendrocyte glycoprotein, depicted in Fig. 17 (SEQ ID NO: \_\_\_\_\_).

30 In a preferred embodiment, peptides derived from NS-specific, self antigens or derivatives of NS-specific antigens activate T-cells, but do not induce an autoimmune disease. An

example of such peptide is a peptide comprising amino acids 51-70 of myelin basic protein. SEQ ID NO:\_\_\_\_ (Kamholz et al., 1986, Proc. Natl. Acad. Sci. USA 83:4962-4966, GenBank accession number M13577; Roth et al., 1987, J. Neurosci. Res. 5 17(4):321-328, GenBank accession number M30516).

In addition, a NS-specific antigen may be a crude NS-tissue preparation, e.g., derived from NS tissue obtained from mammalian NS. Such a preparation may include cells, both living or dead cells, membrane fractions of such cells or 10 tissue, etc.

A NS-specific antigen may be obtained by a NS biopsy or necropsy from a mammal including, but not limited to, from a site of CNS injury; from cadavers; from cell lines grown in culture. Additionally, a NS-specific antigen may be a protein 15 obtained by genetic engineering, chemically synthesized, etc.

In addition to NS-specific antigens, the invention also relates to peptides derived from NS-specific antigens or derivatives including chemical derivatives and analogs of NS-specific antigens which are functionally active, i.e., they are 20 capable of displaying one or more known functional activities associated with a full-length NS-specific antigen. Such functional activities include but are not limited to antigenicity [ability to bind (or compete with a CNS-antigen for binding) to an anti-NS-specific antibody], immunogenicity 25 (ability to generate antibody which binds to a NS-specific protein), and ability to interact with T-cells, resulting in activation comparable to that obtained using the corresponding full-length antigen.

A peptide derived from a CNS-specific or PNS-specific 30 antigen has a sequence comprised within the antigen sequence and is either: (1) an immunogenic peptide, i.e., a peptide that can elicit a human T-cell response detected by T-cell

proliferation or by cytokine (e.g. interferon (IFN)- $\gamma$ , interleukin (IL)-2, IL-4 or IL-10) production or (2) a "cryptic epitope" (also designated herein as "immunosilent" or "nonimmunodominant" epitope), i.e., a peptide that by itself 5 can induce a T-cell immune response that is not induced by the whole antigen protein (see Moalem et al., 1999, *Nature Med.* 5(1)). Cryptic epitopes for use in the present invention include, but are not limited to, peptides of the myelin basic protein sequence: peptide p11-30, p51-70, p91-110, p131-150, 10 and p151-170. Other peptides can be identified by their capacity to elicit a human T-cell response detected by T-cell proliferation or by cytokine (e.g. IFN- $\gamma$ , IL-2, IL-4, or IL-10) production.

In a specific embodiment of the invention, peptides 15 consisting of or comprising a fragment of a NS-specific antigen consisting of at least 10 (contiguous) amino acids of the NS-specific antigen is provided. In other embodiments, the fragment consists of at least 20 contiguous amino acids or 50 contiguous amino acids of the NS-specific antigen.

Derivatives of a NS-specific antigen also include but are not limited to those molecules comprising regions that are substantially homologous to the full-length antigen or fragments thereof (e.g., in various embodiments, at least 60% or 70% or 80% or 90% or 95% identity over an amino acid 25 sequence of identical size or when compared to an aligned sequence in which the alignment is done by a computer homology program known in the art) or whose encoding nucleic acid is capable of hybridizing to a coding nucleotide sequence of the full-length NS-specific antigen, under high stringency, 30 moderate stringency, or low stringency conditions.

Computer programs for determining homology may include but are not limited to TBLASTN, BLASTP, FASTA, TFASTA, and CLUSTALW

(Pearson and Lipman, 1988, Proc. Natl. Acad. Sci. USA 85(8):2444-8; Altschul et al., 1990, J. Mol. Biol. 215(3):403-10; Thompson, et al., 1994, Nucleic Acids Res. 22(22):4673-80; Higgins, et al., 1996, Methods Enzymol 266:383-402; Altschul, 5 et al., 1990, J. Mol. Biol. 215(3):403-10).

The NS-specific antigen derivatives of the invention can be produced by various methods known in the art. The manipulations which result in their production can occur at the gene or protein level. For example, a cloned gene sequence can 10 be modified by any of numerous strategies known in the art (Maniatis, T., 1990, Molecular Cloning, A Laboratory Manual, 2d ed., Cold Spring Harbor Laboratory, Cold Spring Harbor, New York). The sequence can be cleaved at appropriate sites with restriction endonuclease(s), followed by further enzymatic 15 modification if desired, isolated, and ligated *in vitro*.

Additionally, the coding nucleic acid sequence can be mutated *in vitro* or *in vivo*, to create and/or destroy translation, initiation, and/or termination sequences, or to create variations in coding regions and/or form new restriction 20 endonuclease sites or destroy preexisting ones, to facilitate further *in vitro* modification. Any technique for mutagenesis known in the art can be used, including but not limited to, chemical mutagenesis, *in vitro* site-directed mutagenesis (Hutchinson, C., et al., 1978, J. Biol. Chem 253:6551), etc.

25 Manipulations may also be made at the protein level. Included within the scope of the invention are derivatives which are differentially modified during or after translation, e.g., by glycosylation, acetylation, phosphorylation, amidation, derivatization by known protecting/blocking groups, 30 proteolytic cleavage, linkage to an antibody molecule or other cellular ligand, etc. Any of numerous chemical modifications may be carried out by known techniques, including but not

limited to specific chemical cleavage by cyanogen bromide, trypsin, chymotrypsin, papain, V8 protease, NaBH<sub>4</sub>; acetylation, formylation, oxidation, reduction; metabolic synthesis in the presence of tunicamycin; etc.

5        In addition, derivatives of a NS-specific antigen can be chemically synthesized. For example, a peptide corresponding to a portion of an antigen which comprises the desired domain or which mediates the desired activity can be synthesized by use of a peptide synthesizer. Furthermore, if desired, 10 nonclassical amino acids or chemical amino acid analogs can be introduced as a substitution or addition into the amino acid sequence. Non-classical amino acids include but are not limited to the D-isomers of the common amino acids,  $\alpha$ -amino isobutyric acid, 4-aminobutyric acid, Abu, 2-amino butyric 15 acid,  $\gamma$ -Abu,  $\epsilon$ -Ahx, 6-amino hexanoic acid, Aib, 2-amino isobutyric acid, 3-amino propionic acid, ornithine, norleucine, norvaline, hydroxyproline, sarcosine, citrulline, cysteic acid, t-butylglycine, t-butylalanine, phenylglycine, cyclohexylalanine,  $\beta$ -alanine, fluoro-amino acids, designer 20 amino acids such as  $\beta$ -methyl amino acids, C $\alpha$ -methyl amino acids, N $\alpha$ -methyl amino acids, and amino acid analogs in general. Furthermore, the amino acid can be D (dextrorotary) or L (levorotary).

      The functional activity of NS-specific antigens and 25 peptides derived therefrom and derivatives thereof can be assayed by various methods known in the art, including, but not limited to T-cell proliferation assays (Mor and Cohen, 1995, J. Immunol. 155:3693-3699).

      A NS-specific antigen or peptide derived therefrom or 30 derivative thereof may be kept in solution or may be provided in a dry form, e.g. as a powder or lyophilizate, to be mixed with appropriate solution prior to use.

**5.3 NUCLEOTIDE SEQUENCES ENCODING  
NS-ANTIGENS AND PEPTIDES DERIVED THEREFROM**

Compositions comprising a nucleotide sequence encoding a NS-specific antigen or peptide derived therefrom can be used for preventing or inhibiting the effects of injury or disease that result in CNS or PNS degeneration or for promoting nerve regeneration in the CNS or PNS. Specific illustrative examples of useful nucleotide sequences encoding NS-specific antigens or peptides derived from a NS-specific antigen, include but are not limited to nucleotide sequences encoding rat myelin basic protein (MBP) peptides, depicted in Fig. 9 (SEQ ID NO: \_\_\_\_\_); human MBP, depicted in Fig. 10 (SEQ ID NO: \_\_\_\_\_); human myelin PLP, depicted in Figs. 11(A-F) (SEQ ID NO: \_\_\_\_\_); human MOG, depicted in Fig. 12 (SEQ ID NO: \_\_\_\_\_); rat PLP and variant, depicted in Fig. 13 (SEQ ID NO: \_\_\_\_\_); and rat MAG, depicted in Fig. 14 (SEQ ID NO: \_\_\_\_\_).

**5.4 THERAPEUTIC USES**

The compositions described in Sections 5.1 through 5.3 may be used to promote nerve regeneration or to prevent or inhibit secondary degeneration which may otherwise follow primary NS injury, e.g. blunt trauma, penetrating trauma, hemorrhagic stroke, ischemic stroke or damages caused by surgery such as tumor excision. In addition, such compositions may be used to ameliorate the effects of disease that result in a degenerative process, e.g. degeneration occurring in either grey or white matter (or both) as a result of various diseases or disorders which are not recognized by those of reasonable skill in the art as being autoimmune diseases or disorders including, without limitation: Diabetic neuropathy, senile dementias, Alzheimer's disease, Parkinson's Disease, facial nerve (Bell's) palsy, glaucoma, Huntington's chorea, amyotrophic lateral sclerosis (ALS), non-arteritic optic neuropathy, intervertebral

disc herniation, vitamin deficiency, prion diseases such as Creutzfeldt-Jakob disease, carpal tunnel syndrome, peripheral neuropathies associated with various diseases, including but not limited to, uremia, porphyria, hypoglycemia, Sjögren-  
5 Larsson syndrome, acute sensory neuropathy, chronic ataxic neuropathy, biliary cirrhosis, primary amyloidosis, obstructive lung diseases, acromegaly, malabsorption syndromes, polycythemia vera, IgA and IgG gammopathies, complications of various drugs (e.g. metronidazole) and toxins (e.g. alcohol or  
10 organophosphates), Charcot-Marie-Tooth disease, ataxia telangiectasia, Friedreich's ataxia, amyloid polyneuropathies, adrenomyeloneuropathy, Giant axonal neuropathy, Refsum's disease, Fabry's disease, lipoproteinemia, etc.

In a preferred embodiment, the NS-specific antiself  
15 activated T-cells, the NS-specific antigens, peptides derived therefrom, derivatives thereof or the nucleotides encoding said antigens, or peptides or any combination thereof of the present invention are used to treat diseases or disorders which are not autoimmune diseases or neoplasias. In a preferred embodiment,  
20 the compositions of the present invention are administered to a human subject.

#### **5.5 FORMULATIONS AND ADMINISTRATION**

Pharmaceutical compositions for use in accordance with the  
25 present invention may be formulated in conventional manner using one or more physiologically acceptable carriers or excipients. The carrier(s) must be "acceptable" in the sense of being compatible with the other ingredients of the composition and not deleterious to the recipient thereof.

30 The term "carrier" refers to a diluent, adjuvant, excipient, or vehicle with which the therapeutic is administered. The carriers in the pharmaceutical composition

may comprise a binder, such as microcrystalline cellulose, polyvinylpyrrolidone (polyvidone or povidone), gum tragacanth, gelatine, starch, lactose or lactose monohydrate; a disintegrating agent, such as alginic acid, maize starch and 5 the like; a lubricant or surfactant, such as magnesium stearate, or sodium lauryl sulphate; a glidant, such as colloidal silicon dioxide; a sweetening agent, such as sucrose or saccharin; and/or a flavoring agent, such as peppermint, methyl salicylate, or orange flavoring.

10 Methods of administration include, but are not limited to, parenteral, e.g. intravenous, intraperitoneal, intramuscular, subcutaneous, mucosal (e.g., oral, intranasal, buccal, vaginal, rectal, intraocular), intrathecal and intradermal routes. Administration can be systemic or local.

15 For oral administration, the pharmaceutical preparation may be in liquid form, for example, solutions, syrups or suspensions, or may be presented as a drug product for reconstitution with water or other suitable vehicle before use. Such liquid preparations may be prepared by conventional means 20 with pharmaceutically acceptable additives such as suspending agents (e.g., sorbitol syrup, cellulose derivatives or hydrogenated edible fats); emulsifying agents (e.g., lecithin or acacia); non-aqueous vehicles (e.g., almond oil, oily esters, or fractionated vegetable oils); and preservatives 25 (e.g., methyl or propyl-p-hydroxybenzoates or sorbic acid). The pharmaceutical compositions may take the form of, for example, tablets or capsules prepared by conventional means with pharmaceutically acceptable excipients such as binding agents (e.g., pregelatinized maize starch, polyvinyl 30 pyrrolidone or hydroxypropyl methylcellulose); fillers (e.g., lactose, microcrystalline cellulose or calcium hydrogen phosphate); lubricants (e.g., magnesium stearate, talc or

silica); disintegrants (e.g., potato starch or sodium starch glycolate); or wetting agents (e.g., sodium lauryl sulphate). The tablets may be coated by methods well-known in the art.

Preparations for oral administration may be suitably 5 formulated to give controlled release of the active compound.

For buccal administration, the compositions may take the form of tablets or lozenges formulated in conventional manner.

The compositions may be formulated for parenteral administration by injection, e.g., by bolus injection or 10 continuous infusion. Formulations for injection may be presented in unit dosage form, e.g., in ampoules or in multi-dose containers, with an added preservative. The compositions may take such forms as suspensions, solutions or emulsions in oily or aqueous vehicles, and may contain formulatory agents 15 such as suspending, stabilizing and/or dispersing agents. Alternatively, the active ingredient may be in powder form for constitution with a suitable vehicle, e.g., sterile pyrogen-free water, before use.

The compositions may also be formulated in rectal 20 compositions such as suppositories or retention enemas, e.g., containing conventional suppository bases such as cocoa butter or other glycerides.

For administration by inhalation, the compositions for use according to the present invention are conveniently delivered 25 in the form of an aerosol spray presentation from pressurized packs or a nebulizer, with the use of a suitable propellant, e.g., dichlorodifluoromethane, trichlorofluoromethane, dichlorotetrafluoroethane, carbon dioxide or other suitable gas. In the case of a pressurized aerosol the dosage unit may 30 be determined by providing a valve to deliver a metered amount. Capsules and cartridges of, e.g., gelatin for use in an inhaler or insufflator may be formulated containing a powder mix of the

compound and a suitable powder base such as lactose or starch.

In a preferred embodiment, compositions comprising NS-specific antiself activated T-cells, a NS-specific antigen or peptide derived therefrom, or derivative thereof, or a 5 nucleotide sequence encoding such antigen or peptide are formulated in accordance with routine procedures as pharmaceutical compositions adapted for intravenous or intraperitoneal administration to human beings. Typically, compositions for intravenous administration are solutions in 10 sterile isotonic aqueous buffer. Where necessary, the composition may also include a solubilizing agent and a local anesthetic such as lignocaine to ease pain at the site of the injection. Generally, the ingredients are supplied either separately or mixed together. Where the composition is to be 15 administered by infusion, it can be dispensed with an infusion bottle containing sterile pharmaceutical grade water or saline. Where the composition is administered by injection, an ampoule of sterile water or saline for injection can be provided so that the ingredients may be mixed prior to administration.

20 Pharmaceutical compositions comprising NS-specific antigen or peptide derived therefrom or derivative thereof may optionally be administered with an adjuvant, such as Incomplete Freund's Adjuvant.

The invention also provides a pharmaceutical pack or kit 25 comprising one or more containers filled with one or more of the ingredients of the pharmaceutical compositions of the invention.

In a preferred embodiment, the pharmaceutical compositions of the invention are administered to a mammal, preferably a 30 human, shortly after injury or detection of a degenerative lesion in the NS. The therapeutic methods of the invention may comprise administration of a NS-specific antiself activated T-

cell, or a NS-specific antigen or peptide derived therefrom or derivative thereof, or a nucleotide sequence encoding such antigen or peptide or any combination thereof. The NS-specific antigen may be administered before, concurrently or after 5 administration of NS-specific antiself activated T-cells, a peptide derived from a NS-specific antigen or derivative thereof or a nucleotide sequence encoding such antigen or peptide.

In an embodiment, the compositions of the invention are 10 administered in combination with one or more of the following:

(a) mononuclear phagocytes, preferably cultured monocytes (as described in PCT publication No. WO 97/09985, which is incorporated herein by reference in its entirety), that have been stimulated to enhance their capacity to promote axonal 15 regeneration; (b) a neurotrophic factor such as acidic fibroblast growth factor; and (c) an anti-inflammatory therapeutic substance (i.e., an anti-inflammatory steroid, such as dexamethasone or methylprednisolone, or a non-steroidal anti-inflammatory agent or drug, such as aspirin, indomethacin, 20 ibuprofen, fenoprofen, ketoprofen or haproxen, or an anti-inflammatory peptide, such as Thr-Lys-Pro (TKP)).

In an embodiment, mononuclear phagocyte cells according PCT Publication No. WO 97/09985 and U.S. patent application Serial No. 09/041,280, filed March 11, 1998, are injected into 25 the site of injury or lesion within the CNS, either concurrently, prior to, or following parenteral administration of NS-specific antiself activated T-cells, a NS-specific antigen or peptide derived therefrom or derivative thereof, or a nucleotide sequence encoding such antigen or peptide.

In an embodiment, administration of NS-specific activated 30 T-cells, a NS-specific antigen or peptide derived therefrom or derivative thereof, or a nucleotide sequence encoding such antigen or peptide, may be administered as a single dose or may

be repeated, preferably at 2 week intervals and then successively longer intervals once a month, once a quarter, once every six months, etc. The course of treatment may last several months, several years or occasionally also through the 5 life-time of the individual, depending on the condition or disease which is being treated. In the case of a CNS injury, the treatment may range between several days to months or even years, until the condition has stabilized and there is no or only a limited risk of development of secondary degeneration. In chronic human diseases or conditions such as Alzheimer's 10 disease or Parkinson's disease, the therapeutic treatment in accordance with the invention may be for life.

As will be evident to those of skill in the art, the therapeutic effect depends at times on the condition or disease to be treated, on the individual's age and health condition, on 15 other physical parameters (e.g. gender, weight, etc.) of the individual, as well as on various other factors, e.g. whether the individual is taking other drugs, etc.

The optimal dose of the therapeutic compositions comprising NS-specific antiself activated T-cells of the 20 invention is proportional to the number of nerve fibers affected by CNS injury or disease at the site being treated. In a preferred embodiment, the dose ranges from about  $5 \times 10^6$  to about  $10^7$  for treating a lesion affecting about  $10^5$  nerve fibers, such as a complete transection of a rat optic nerve, and ranges from about  $10^7$  to about  $10^8$  for treating a lesion 25 affecting about  $10^6$  -  $10^7$  nerve fibers, such as a complete transection of a human optic nerve. As will be evident to those of skill in the art, the dose of T-cells can be scaled up or down in proportion to the number of nerve fibers thought to be affected at the lesion or site of injury being treated.

30 The following examples illustrate certain features of the present invention but are not intended to limit the scope of the present invention.

6. EXAMPLE: ACCUMULATION OF ACTIVATED  
T-CELLS IN INJURED OPTIC NERVE

5 6.1 MATERIALS AND METHODS

6.1.1 ANIMALS

Female Lewis rats were supplied by the Animal Breeding Center of the Weizmann Institute of Science (Rehovot, IL), matched for age (8-12 weeks) and housed four to a cage in a 10 light and temperature-controlled room.

6.1.2 MEDIA

The T-cell proliferation medium contained the following: Dulbecco's modified Eagle's medium (DMEM, Biological 15 Industries, Israel) supplemented with 2mM L-glutamine (L-Glu, Sigma, USA),  $5 \times 10^{-5}$ M 2-mercaptoethanol (2-ME, Sigma), penicillin (100 IU/ml; Biological Industries), streptomycin (100  $\mu$ g/ml; Biological Industries), sodium pyruvate (1 mM; Biological Industries), non-essential amino acids (1 ml/100 ml; Biological Industries) and autologous rat serum 1% (vol/vol) (Mor et al., 20 Clin. Invest., 85:1594 (1990)). Propagation medium contained: DMEM, 2-ME, L-Glu, sodium pyruvate, non-essential amino acids and antibiotics in the same concentration as above with the addition of 10% fetal calf serum (FCS), and 10% T-cell growth factor (TCGF) obtained from the supernatant of concanavalin A- 25 stimulated spleen cells (Mor et al., *supra*, 1990).

6.1.3 ANTIGENS

Myelin basic protein (MBP) from the spinal cords of guinea pigs was prepared as described (Hirshfeld, et al., 1970, FEBS 30 Lett. 7:317). Ovalbumin was purchased from Sigma (St. Louis, Missouri). The p51-70 of the rat 18.5kDa isoform of MBP (sequence: APKRGSGKDSHRTTHYG) SEQ ID NO: \_\_\_\_\_ and the p277

peptide of the human hsp60 (sequence: VLGGGCALLRCPALDSLTPANED) SEQ ID NO: \_\_\_\_ (Elias, et al., 1991, Proc. Natl. Acad. Sci. USA 88, 3088-91) were synthesized using the 9-fluorenylmethoxycarbonyl technique with an automatic multiple 5 peptide synthesizer (AMS 422, ABIMED, Langenfeld, Germany). The purity of the peptides was analyzed by HPLC and amino acid composition.

#### **6.1.4 T-CELL LINES**

10 T-cell lines were generated from draining lymph node cells obtained from Lewis rats immunized with an antigen (described above in Section 6.1.3). The antigen was dissolved in PBS (1mg/ml) and emulsified with an equal volume of incomplete Freund's adjuvant (Difco Laboratories, Detroit, Michigan) supplemented with 4 mg/ml *Mycobacterium tuberculosis* (Difco 15 Laboratories, Detroit, Michigan). The emulsion (0.1 ml) was injected into hind foot pads of the rats. Ten days after the antigen was injected, the rats were killed and draining lymph nodes were surgically removed and dissociated. The cells were washed and activated with the antigen (10 $\mu$ g/ml) in 20 proliferation medium (described above in Section 6.1.2). After incubation for 72 h at 37°C, 90% relative humidity and 7% CO<sub>2</sub>, the cells were transferred to propagation medium (described above in Section 6.1.2). Cells were grown in propagation medium for 4-10 days before being re-exposed to antigen 25 (10 $\mu$ g/ml) in the presence of irradiated (2000 rad) thymus cells (10<sup>7</sup> cells/ml) in proliferation medium. The T-cell lines were expanded by repeated re-exposure and propagation.

#### **6.1.5 CRUSH INJURY OF RAT OPTIC NERVE**

30 Crush injury of the optic nerve was performed as previously described (Duvdevani et al., 1990, Neurol. Neurosci. 2:31-38). Briefly, rats were deeply anesthetized by i.p.

injection of Rompun (xylazine, 10 mg/kg; Vitamed, Israel) and Vetalar (ketamine, 50 mg/kg; Fort Dodge Laboratories, Fort Dodge, Iowa). Using a binocular operating microscope, a lateral canthotomy was performed in the right eye and the conjunctiva was incised lateral to the cornea. After separation of the retractor bulbi muscles, the optic nerve was exposed intraorbitally by blunt dissection. Using calibrated cross-action forceps, a moderate crush injury was inflicted on the optic nerve, 2 mm from the eye (Duvdevani et al., 10 *Instructure Neurology and Neuroscience*, 2:31, 1990). The contralateral nerve was left undisturbed and was used as a control.

#### 6.1.6 IMMUNOCYTOCHEMISTRY OF T-CELLS

15 Longitudinal cryostat nerve sections (20  $\mu\text{m}$  thick) were picked up onto gelatin glass slides and frozen until preparation for fluorescent staining. Sections were thawed and fixed in ethanol for 10 minutes at room temperature, washed twice with double-distilled water (ddH<sub>2</sub>O), and incubated for 3 20 minutes in PBS containing 0.05% polyoxyethylene-sorbitan monolaurate (Tween-20; Sigma, USA). Sections were then incubated for 1 hr at room temperature with a mouse monoclonal antibody directed against rat T-cell receptor (TCR) (1:100, Hunig et al., *J. Exp. Med.*, 169:73, 1989), in PBS containing 3% 25 FCS and 2% BSA. After three washes with PBS containing 0.05% Tween-20, the sections were incubated with fluorescein isothiocyanate-conjugated goat anti-mouse IgG (with minimal cross-reaction to rat, human, bovine and horse serum proteins) (Jackson ImmunoResearch, West Grove, Pennsylvania) for 1 hr at 30 room temperature. The sections were then washed with PBS containing Tween-20 and treated with glycerol containing 1,4-diazobicyclo-(2,2,2) octane (Sigma), to inhibit quenching of fluorescence. The sections were viewed with a Zeiss microscope

and cells were counted. Staining in the absence of first antibody was negative.

5        **6.2 RESULTS**

Fig. 1 shows accumulation of T-cells measured immuno-histochemically. The number of T cells was considerably higher in injured nerves of rats injected with anti-MBP, anti-OVA or anti-p277 cells; statistical analysis (one-way ANOVA) showed significant differences between T cell numbers in injured optic 10 nerves of rats injected with anti-MBP, anti-OVA, or anti-p277 T cells and in injured optic nerves of rats injected with PBS ( $P<0.001$ ); and between injured optic nerves and uninjured optic nerves of rats injected with anti-MBP, anti-OVA, or anti-p277 T cells ( $P<0.001$ ).  
15

7. **EXAMPLE: NEUROPROTECTION BY AUTOIMMUNE ANTI-MBP T-CELLS**

7.1 **MATERIALS AND METHODS**

Animals, media, antigens, crush injury of rat optic nerve, sectioning of nerves, T-cell lines, and immunolabeling of nerve 20 sections are described in Section 6, *supra*.

7.1.1 **RETROGRADE LABELING AND MEASUREMENT OF PRIMARY DAMAGE AND SECONDARY DEGENERATION**

Primary damage of the optic nerve axons and their attached 25 retinal ganglion cells (RGCs) were measured after the immediate post-injury application of the fluorescent lipophilic dye 4-(4-(didecylamino)styryl)-n-methylpyridinium iodide (4-Di-10-Asp) (Molecular Probes Europe BV, Netherland) distal to the site of injury. Only axons that are intact are capable of transporting the dye back to their cell bodies; therefore, the number of 30 labeled cell bodies is a measure of the number of axons that survived the primary damage. Secondary degeneration was also measured by application of the dye distal to the injury site,

but 2 weeks after the primary lesion was inflicted. Application of the neurotracer dye distal to the site of the primary crush after 2 weeks ensures that only axons that survived both the primary damage and the secondary degeneration 5 will be counted. This approach makes it possible to differentiate between neurons that are still functionally intact and neurons in which the axons are injured but the cell bodies are still viable, as only those neurons whose fibers are morphologically intact can take up dye applied distally to the site of injury and transport it to their cell bodies. Using 10 this method, the number of labeled ganglion cells reliably reflects the number of still-functioning neurons. Labeling and measurement were done by exposing the right optic nerve for a second time, again without damaging the retinal blood supply. Complete axotomy was done 1-2 mm from the distal border of the 15 injury site and solid crystals (0.2-0.4 mm in diameter) of 4-Di-10-Asp were deposited at the site of the newly formed axotomy. Uninjured optic nerves were similarly labeled at approximately the same distance from the globe. Five days after dye application, the rats were killed. The retina was 20 detached from the eye, prepared as a flattened whole mount in 4% paraformaldehyde solution and examined for labeled ganglion cells by fluorescence microscopy. The percentage of RGCs surviving secondary degeneration was calculated using the following formula: (Number of spared neurons after secondary degeneration) / (Number of spared neurons after primary damage) 25 x 100.

#### **7.1.2 ELECTROPHYSIOLOGICAL RECORDINGS**

Nerves were excised and their compound action potentials (CAPs) were recorded *in vitro* using a suction 30 electrode experimental set-up (Yoles, E. et al., 1996, *J. Neurotrauma*, 13:49-57). At different times after injury and injection of T cells or PBS, rats were killed by

intraperitoneal injection of pentobarbitone (170 mg/kg) (CTS Chemical Industries, Israel). Both optic nerves were removed while still attached to the optic chiasma, and were immediately transferred to a vial containing a fresh salt solution 5 consisting of 126 mM NaCl, 3 mM KCl, 1.25 mM NaH<sub>2</sub>PO<sub>4</sub>, 26 mM NaHCO<sub>3</sub>, 2 mM MgSO<sub>4</sub>, 2 mM CaCl<sub>2</sub> and 10 mM D-glucose, aerated with 95% O<sub>2</sub> and 5% CO<sub>2</sub> at room temperature. After 1 hour, electrophysiological recordings were made. In the injured nerve, recordings were made in a segment distal to the injury site. This segment contains axons of viable retinal ganglion 10 cells that have escaped both primary and secondary damage, as well as the distal stumps of non-viable retinal ganglion cells that have not yet undergone Wallerian degeneration. The nerve ends were connected to two suction Ag-AgCl electrodes immersed in the bathing solution at 37°C. A stimulating pulse was 15 applied through the electrode, and the CAP was recorded by the distal electrode. A stimulator (SD9; Grass Medical Instruments, Quincy, Massachusetts) was used for supramaximal electrical stimulation at a rate of 1 pps to ensure stimulation of all propagating axons in the nerve. The measured signal was 20 transmitted to a microelectrode AC amplifier (model 1800; A-M Systems, Everett, Washington). The data were processed using the LabView 2.1.1 data acquisition and management system (National Instruments, Austin, Texas). For each nerve, the difference between the peak amplitude and the mean plateau of eight CAPs was computed and was considered as proportional to 25 the number of propagating axons in the optic nerve. The experiments were done by experimentors 'blinded' to sample identity. In each experiment the data were normalized relative to the mean CAP of the uninjured nerves from PBS-injected rats.

### **7.1.3 CLINICAL EVALUATION OF EXPERIMENTAL AUTOIMMUNE ENCEPHALOMYELITIS**

Clinical disease was scored every 1 to 2 days according to the following neurological scale: 0, no abnormality; 1, tail atony; 2, hind limb paralysis; 3, paralysis extending to thoracic spine; 4, front limb paralysis; 5, moribund state.

## **7.2 RESULTS**

### **7.2.1 NEUROPROTECTION BY AUTOIMMUNE anti-MBP T-CELLS**

10 Morphological analyses were done to assess the effect of the T cells on the response of the nerve to injury, and specifically on secondary degeneration. Rats were injected intraperitoneally immediately after optic nerve injury with PBS or with  $1 \times 10^7$  activated T cells of the various cell lines.

15 The degree of primary damage to the optic nerve axons and their attached RGCs was measured by injecting the dye 4-Di-10-Asp distal to the site of the lesion immediately after the injury. A time lapse of 2 weeks between a moderate crush injury and dye application is optimal for demonstrating the number of still-viable labeled neurons as a measure of secondary degeneration.

20 Therefore, secondary degeneration was quantified by injecting the dye immediately or 2 weeks after the primary injury, and calculating the additional loss of RGCs between the first and the second injections of the dye. The percentage of RGCs that

25 had survived secondary degeneration was then calculated. The percentage of labeled RGCs (reflecting still-viable axons) was significantly greater in the retinas of the rats injected with anti-MBP T cells than in the retinas of the PBS-injected control rats (Fig. 2). In contrast, the percentage of labeled

30 RGCs in the retinas of the rats injected with anti-OVA or anti-p277 T cells was not significantly greater than that in the control retinas. Thus, although the three T-cell lines

accumulated at the site of injury, only the MBP-specific autoimmune T cells had a substantial effect in limiting the extent of secondary degeneration. Labeled RGCs of injured optic nerves of rats injected with PBS (Fig. 3A), with anti-  
5 p277 T cells (Fig. 3B) or with anti-MBP T cells were compared morphologically using micrographs (Fig. 3C).

#### **7.2.2 CLINICAL SEVERITY OF EAE**

10 Animals were injected i.p. with  $10^7$   $T_{MBP}$  cells with or without concurrent optic nerve crush injury. The clinical course of the rats injected with the  $T_{MBP}$  cells was evaluated according to the neurological paralysis scale. Each group contained 5-9 rats. The functional autoimmunity of the injected anti-MBP T-cells was demonstrated by the development of transient EAE in the recipients of these cells. As can be  
15 seen in Fig. 4A, the course and severity of the EAE was not affected by the presence of the optic nerve crush injury.

#### **7.2.3 SURVIVAL OF RGCS IN NON-INJURED NERVES**

20 Animals were injected i.p. with  $10^7$   $T_{MBP}$  cells or PBS. Two weeks later, 4-Di-10-Asp was applied to the optic nerves. After five days the retinas were excised and flat mounted. Labeled RGCs from five fields (located at approximately the same distance from the optic disk), in each retina were counted and their average number per area ( $mm^2$ ) was calculated.  
25

As can be seen in Fig. 4B, there is no difference in the number of surviving RGCs per area ( $mm^2$ ) in non-injured optic nerves of rats injected with anti-MBP T-cells compared to in rats injected with PBS.

#### **7.2.4 NEUROPROTECTION BY T-CELLS REACTIVE TO A CRYPTIC EPITOPE**

30 To determine whether the neuroprotective effect of the anti-MBP T cells is correlated with their virulence, the effect

of T cells reactive to a 'cryptic' epitope of MBP, the peptide 51-70 (p51-70) was examined. 'Cryptic' epitopes activate specific T cells after an animal is immunized with the particular peptide, but not with the whole antigen (Mor, P. et al., 1995, *J. Immunol.* **155**:3693-3699). The T-cell line reactive to the whole MBP and the T-cell line reactive to the cryptic epitope p51-70 were compared for the severity of the EAE they induced, and for their effects on secondary degeneration. In rats injected with the T-cell line reactive to the cryptic epitope, disease severity (as manifested by the maximal EAE score) was significantly lower than that in rats injected with the T-cell line reactive to the whole protein (Table 1). Whereas anti-MBP T cells caused clinical paralysis of the limbs, rats injected with the anti-p51-70 T cells developed only tail atony, not hind limb paralysis, and almost none showed weakness of the hind limbs. Despite this difference in EAE severity, the neuroprotective effect of the less virulent (anti-p51-70) T cells was similar to that of the more virulent (anti-MBP) T cells (Fig. 5). The percentage of RGCs surviving secondary degeneration in the retinas of rats injected with either of the lines was significantly higher than in the retinas of the PBS-injected rats. Thus, there was no correlation between the neuroprotective effect of the autoimmune T cells and their virulence. It is possible that the anti-p51-70 T cells encounter little antigen in the intact CNS, and therefore cause only mild EAE. Their target antigen may however become more available after injury, enabling these T cells to exert a neuroprotective effect.

TABLE 1. Anti-MBP and anti-p51-70 T cells vary in pathogenicity

T cell line	Clinical EAE	Mean max. score
30 Whole MBP	Moderate to severe	2.00 ± 0.25
p51-70 of MBP	Mild	0.70 ± 0.2

Immediately after optic nerve crush injury, Lewis rats were injected with activated anti-MBP T cells or anti-p51-70 T cells. The clinical course of EAE was evaluated according to the neurological paralysis scale. The mean maximal (max.) score  $\pm$  s.e.m. was calculated as the average maximal score of all the diseased rats in each group. The table is 5 a summary of nine experiments. Each group contains five to ten rats. Statistical analysis showed a significant difference between the mean maximal score of rats injected with anti-MBP T cells and that of rats injected with anti-p51-70 T cells ( $P=0.039$ , Student's t-test).

#### 7.2.5 ELECTROPHYSIOLOGICAL ACTIVITY

10 To confirm the neuroprotective effect of the anti-MBP T cells, electrophysiological studies were done. Immediately after optic nerve injury, the rats were injected intraperitoneally with PBS or with  $1 \times 10^7$  activated anti-MBP or anti-OVA T cells. The optic nerves were excised 7, 11 or 14 15 days later and the compound action potentials (CAPs), a measure of nerve conduction, were recorded from the injured nerves. On day 14, the mean CAP amplitudes of the distal segments recorded from the injured nerves obtained from the PBS-injected control rats were 33% to 50% of those recorded from the rats injected 20 with the anti-MBP T cells. (Fig. 6A, Table 2). As the distal segment of the injured nerve contains both axons that escaped the primary insult and injured axons that have not yet degenerated, the observed neuroprotective effect could reflect the rescue of spared neurons, or a delay of Wallerian 25 degeneration of the injured neurons (which normally occurs in the distal stump), or both. No effect of the injected anti-MBP T cells on the mean CAP amplitudes of uninjured nerves was observed (Fig. 6B, Table 2). It is unlikely that the neuroprotective effect observed on day 14 could have been due to the regrowth of nerve fibers, as the time period was too 30 short for this.

The strong neuroprotective effect of the anti-MBP T cells seen on day 14 was associated with a significantly decreased

CAP amplitude recorded on day 7 (Table 2). The anti-MBP T cells manifested no substantial effect on the uninjured nerve on day 7, indicating that the reduction in electrophysiological activity observed in the injured nerve on day 7 might reflect 5 the larger number of T cells present at the injury site relative to the uninjured nerve (Fig. 1). The observed reduction in CAP amplitude in the injured nerve on day 7 reflected a transient reduction in conduction, which may have imposed a transient resting state in the injured nerve. This transient effect had not only disappeared, but was even 10 reversed by day 14 (Table 2). Early signs of the neuroprotective effect could already be detected on day 11 in the rats injected with anti-MBP T cells (data not shown). In rats injected with anti-OVA T cells, no reduction in CAP amplitude on day 7 could be detected in either the injured or 15 the uninjured nerves, and no neuroprotective effect was observed on day 14 (Table 2). Thus, it seems that the early reduction in CAP and the late neuroprotection shown specifically by the anti-MBP T cells are related.

20 TABLE 2. Transient reduction in electrophysiological activity of the injured optic nerve induced by anti-MBP T cells, followed by a neuroprotective effect

	Uninjured optic nerve		Injured optic nerve		
	Day 7	Day 14	Day 7	Day 14	
25	Ratio (%) T <sub>MBP</sub> /PBS	89.9±9.4 (n=22)	101.2±22.7 (n=10)	63.8*±14.9 (n=17)	243.1**±70.8 (n=8)
	Ratio (%) T <sub>OVA</sub> /PBS	109.7±13.2 (n=11)	92.5±12.6 (n=3)	125.5±24.4 (n=11)	107.3±38.9 (n=4)

Immediately after optic nerve injury, rats were injected with PBS or with activated anti-MBP or anti-OVA T cells. After 7 or 14 days, the CAPs of injured and uninjured nerves were recorded. Ratios were calculated for uninjured nerves as (mean CAP of uninjured nerves from T cell-injected rats/mean CAP of uninjured nerves from PBS-injected rats) x 5 100, or for injured nerves as (mean CAP of injured nerves from T cell-injected rats/mean CAP of injured nerves from PBS-injected rats) x 100. The P value was calculated by comparing the logarithms of the normalized CAP amplitudes of nerves from PBS-injected rats and rats injected with T cells, using the unpaired Student's t-test, \*P<0.05; \*\*P<0.01 n=sample size.

10

## 8. EXAMPLE: NEUROPROTECTIVE EFFECTS OF NS-SPECIFIC ANTIGEN

### 8.1 MATERIALS AND METHODS

Animals, crush injury of rat optic nerve, and retrograde labeling are described above in Sections 6 and 7. A peptide 15 based on amino acids 35-55 of myelin/oligodendrocyte glycoprotein (MOG p35-55) was chemically synthesized at the Weizmann Institute, Israel.

#### 8.1.1 INHIBITION OF SECONDARY DEGENERATION

20 Rats were injected intradermally in the footpads with MOG p35-55 (50 µg/animal) and IFA, or PBS ten days prior to optic nerve crush injury. Retinal ganglion cells were assessed two weeks after injury using retrograde labeling as described above. The number of RGCs in rats injected with PBS or MOG p35-55 was expressed as a percentage of the total number of 25 neurons in rats injected with MOG p35-55 in the absence of crush injury.

### 8.2 RESULTS

As shown in Fig. 7, the number of labeled retinal ganglion 30 cells (indicating viable axons) was about 12.5 fold greater in animals injected with MOG p35-55 compared to in animals receiving PBS.

9. EXAMPLE: NEUROPROTECTIVE EFFECTS OF MBP  
ADMINISTERED ORALLY

9.1 MATERIALS AND METHODS

Animals, crush injury of rat optic nerve, and retrograde  
5 labeling of RGCs are described above in Sections 6 and 7.

9.1.1 INHIBITION OF SECONDARY DEGENERATION

Bovine MBP (Sigma, Israel) (1 mg/dose) was administered to  
rats by gavage using a blunt needle. MBP was administered 5  
times, every third day, beginning 2 weeks prior to optic nerve  
10 crush injury. The number of RGCs in treated animals was  
expressed as a percentage of the total number of neurons in  
animals subjected to optic nerve crush injury but which did not  
receive MBP.

15 9.2 RESULTS

As shown in Fig. 8, the number of labeled RGCs was about  
1.3 fold greater in animals treated with MBP compared to  
untreated animals.

20 10. DISCUSSION OF EXPERIMENTAL RESULTS

The results of the experiments described in Sections 6 and  
7 show that activated T-cells accumulate at a site of injury in  
the CNS. Furthermore, the results also demonstrate that the  
accumulation of T-cells at the site of injury is a non-specific  
process, *i.e.*, T-cells which accumulated at the site of injury  
25 included both T-cells which are activated by exposure to an  
antigen present at the site of injury as well as T-cells which  
are activated by an antigen not normally present in the  
individual.

The results of experiments described in Section 7  
30 demonstrate that the beneficial effects of T-cells in  
ameliorating damage due to injury in the CNS are associated  
with a NS-specific self-antigen as illustrated by MBP. More

specifically, the administration of non-recombinant T-cells which were activated by exposure to an antigen which can cause autoimmune disease ( $T_{MBP}$ ), rather than aggravating the injury, led to a significant degree of protection from secondary 5 degeneration. Thus, activating T-cells by exposure to a fragment of a NS-specific antigen was beneficial in limiting the spread of injury in the CNS. The present findings show that secondary degeneration can be inhibited by the transfer into the individual of non-recombinant T-cells which recognize a NS-specific self antigen which is present at a site of 10 injury. The T-cells may recognize cryptic or non-pathogenic epitopes of NS-self antigens.

In addition, the studies described in Sections 8 and 9 show that activation of T-cells by administering an immunogenic antigen (e.g. MBP) or immunogenic epitope of an antigen (e.g. 15 MOG p35-55), may be used for preventing or inhibiting secondary CNS degeneration following injury.

The present invention is not to be limited in scope by the exemplified embodiments, which are intended as illustrations of 20 single aspects of the invention. Indeed, various modifications of the invention in addition to those shown and described herein will become apparent to those skilled in the art from the foregoing description and accompanying drawings. Such modifications are intended to fall within the scope of the appended claims.

25 All publications cited herein are incorporated by reference in their entirety.

WHAT IS CLAIMED IS:

1. A method for preventing or inhibiting axonal degeneration in the central nervous system or peripheral nervous system comprising administering to a human in need thereof:

- 5 (a) non-recombinant, NS-specific antiself activated T-cells;
- (b) a NS-specific antigen;
- (c) a peptide derived from a NS-specific antigen;
- (d) a nucleotide sequence encoding a NS-specific antigen;
- (e) a nucleotide sequence encoding a peptide derived from a NS-specific antigen; or
- (f) any combination of (a)-(e),

10 to ameliorate the effects of injury or disease.

15

2. A method for promoting nerve regeneration in the central nervous system or peripheral nervous system comprising administering to a human in need thereof:

- 20 (a) non-recombinant, NS-specific antiself activated T-cells;
- (b) a NS-specific antigen;
- (c) a peptide derived from a NS-specific antigen;
- (d) a nucleotide sequence encoding a NS-specific antigen;
- (e) a nucleotide sequence encoding a peptide derived from a NS-specific antigen; or
- (f) any combination of (a)-(e),

25

to ameliorate the effects of injury or disease.

30

3. The method according to claim 1 or 2 in which said injury comprises blunt trauma, penetrating trauma, hemorrhagic stroke, ischemic stroke, or damages caused by

surgery.

4. The method of claim 1 or 2 in which said disease is Diabetic neuropathy, senile dementia, Alzheimer's disease, 5 Parkinson's Disease, facial nerve (Bell's) palsy, glaucoma, Huntington's chorea, amyotrophic lateral sclerosis, non-arteritic optic neuropathy, or vitamin deficiency.

5. The method of claim 1 or 2 in which said disease 10 is not an autoimmune disease or a neoplasm.

6. The method of claim 1 or 2 in which said peptide derived from a NS-specific antigen is an immunogenic epitope or a cryptic epitope.

15

7. The method according to claims 1 or 2 in which said NS-specific antigen is administered intravenously, intraperitoneally, intramuscularly, subcutaneously, orally, intranasally, vaginally, rectally, intraocularly, intrathecally, 20 intradermally, or buccally.

8. The method according to claim 1(a), 1(c), 1(d), 1(e), 2(a), 2(c), 2(d), or 2(e), further comprising administering to a human in need thereof a NS-specific antigen.

25

9. The method according to claim 8 in which said NS-specific antigen is administered before or after administration of the composition according to claim 1(a), 1(c), 1(d), 1(e), 2(a), 2(c) or 2(e).

30

10. The method according to claim 8 in which said NS-specific antigen is administered concurrently with

administration of the composition according to claim 1(a), 1(c), 1(d), 1(e), 2(a), 2(c) or 2(e).

11. The method according to claim 1 or 2 in which 5 said T-cells are attenuated.

12. The method according to claim 1 or 2 in which said T-cells are autologous or allogeneic.

10 13. The method according to claim 1 or 2 in which the NS-specific antigen or peptide derived therefrom is myelin basic protein, myelin oligodendrocyte glycoprotein, proteolipid protein, myelin-associated glycoprotein, S-100,  $\beta$ -amyloid, Thy-1, P0, or P2.

15 14. The method according to claim 1d or 2d in which the nucleotide sequence is depicted in Fig. 9, Fig. 10, Fig. 11(A-F), Fig. 12, Fig. 13, or Fig. 14.

20 15. The method according to claim 1 or 2 in which the NS-specific antigen comprises the amino acid sequence of Fig. 15, Fig. 16, or Fig. 17.

25

30

ABSTRACT OF THE INVENTION

The present invention discloses compositions and methods for the treatment of injury or disease of the nervous system CNS. In a particular embodiment, the invention provides methods 5 of treatment using non-recombinant activated antiself T-cells that recognize an antigen of the NS or a peptide derived therefrom or a derivative thereof to promote nerve regeneration or to prevent or inhibit axonal degeneration within the NS. The invention also provides methods of treatment using a NS-specific 10 antigen or peptide derived therefrom or a derivative thereof or a nucleotide sequence encoding said antigen or peptide to promote nerve regeneration or to prevent or inhibit axonal degeneration in NS, i.e., the CNS and/or PNS. The NS-specific antiself activated T-cells may be administered alone or in 15 combination with NS-specific antigen or peptide derived therefrom or a derivative thereof or a nucleotide sequence encoding said antigen or peptide or any combination thereof.

20

25

30

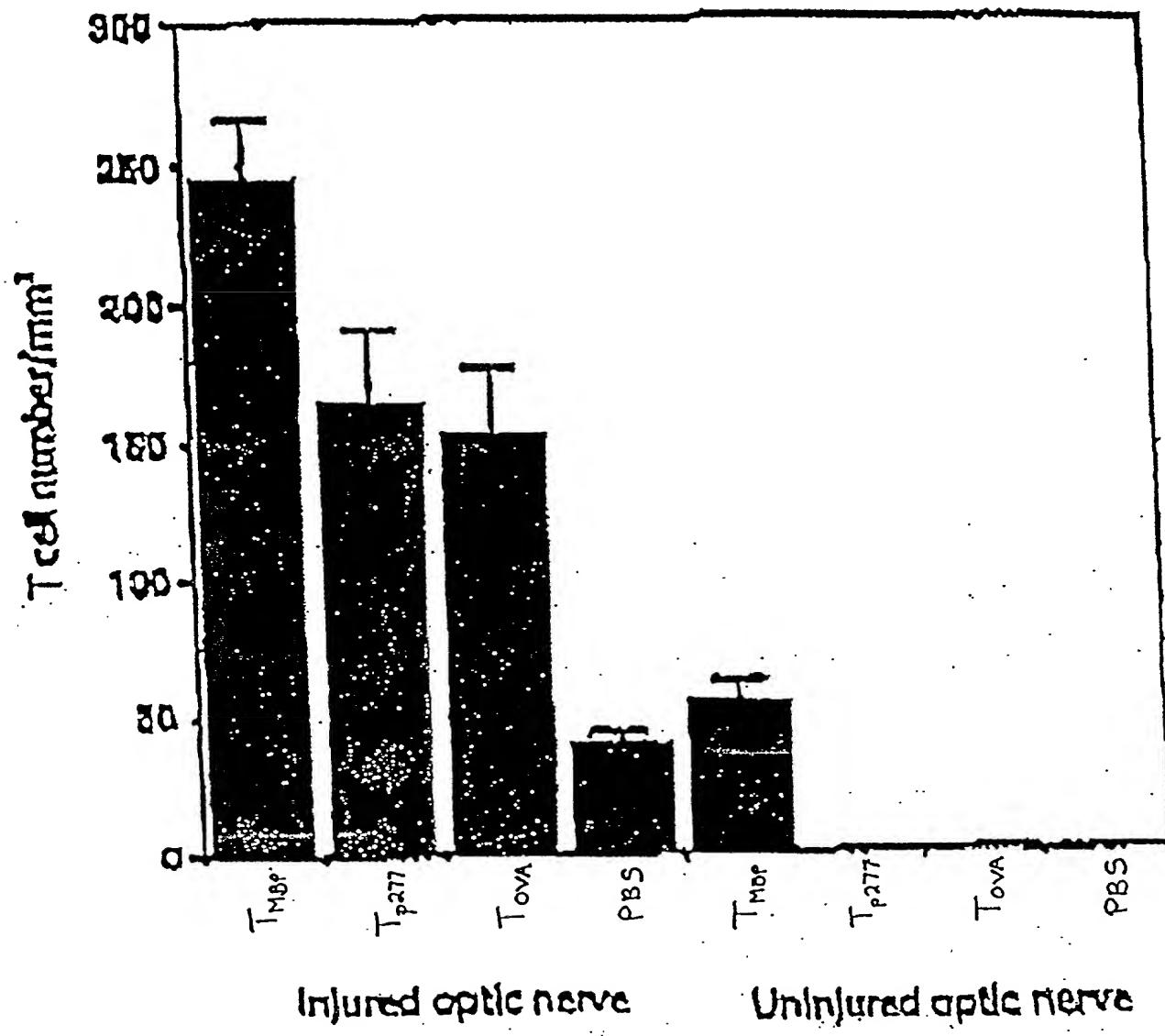


FIG. 1

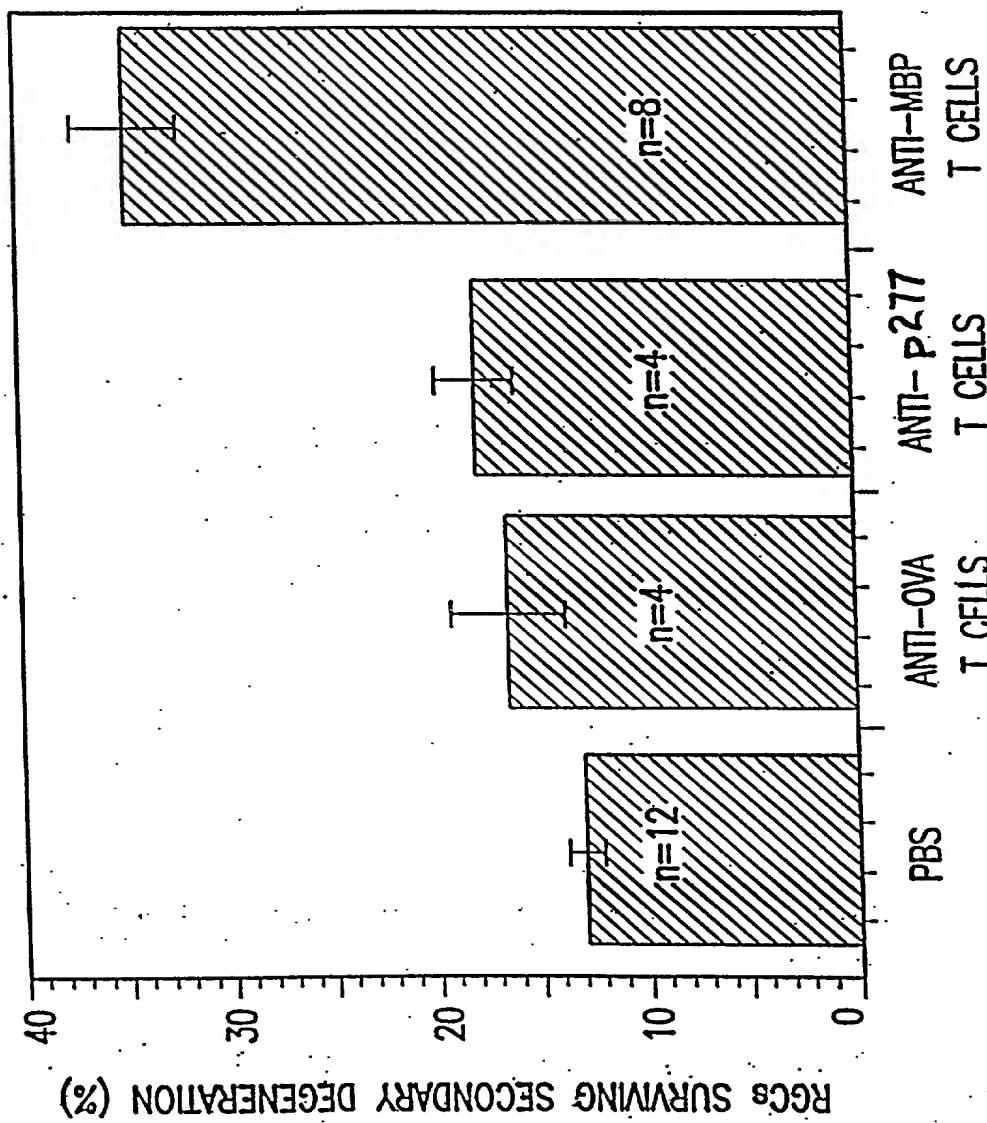
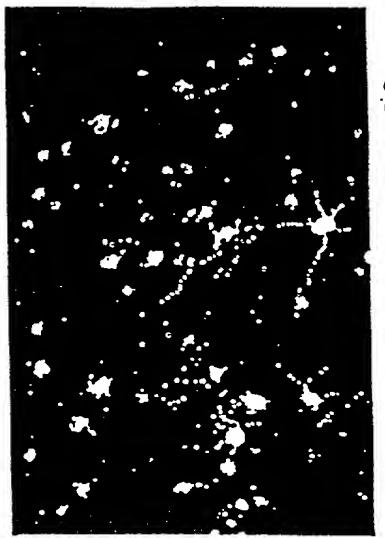


FIG. 2



160  $\mu$ m

FIG. 3C

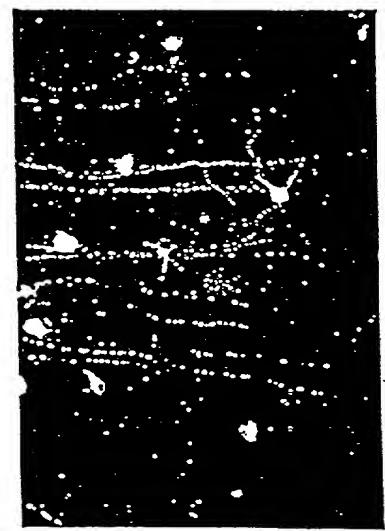


FIG. 3B



FIG. 3A

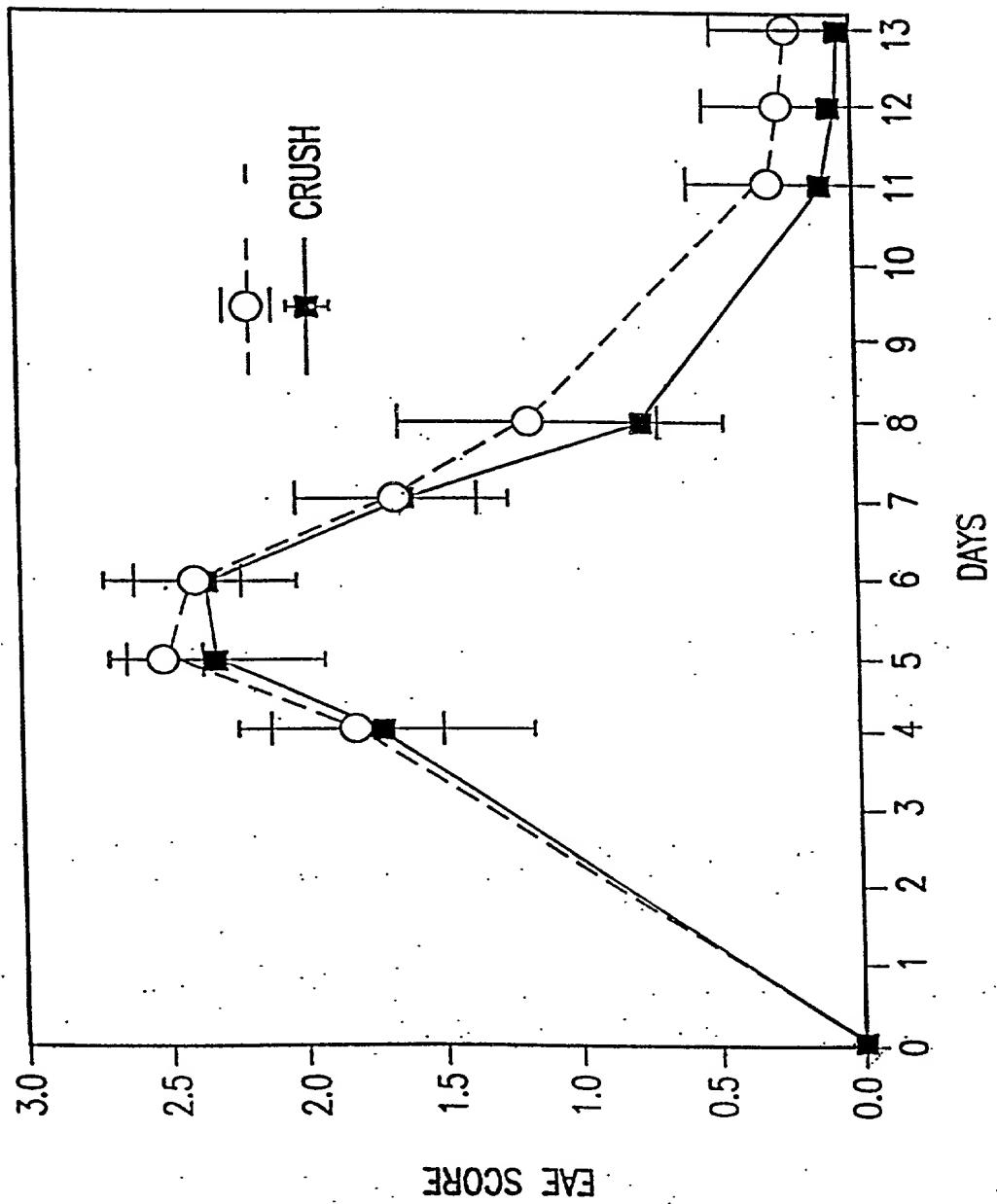


FIG. 4A

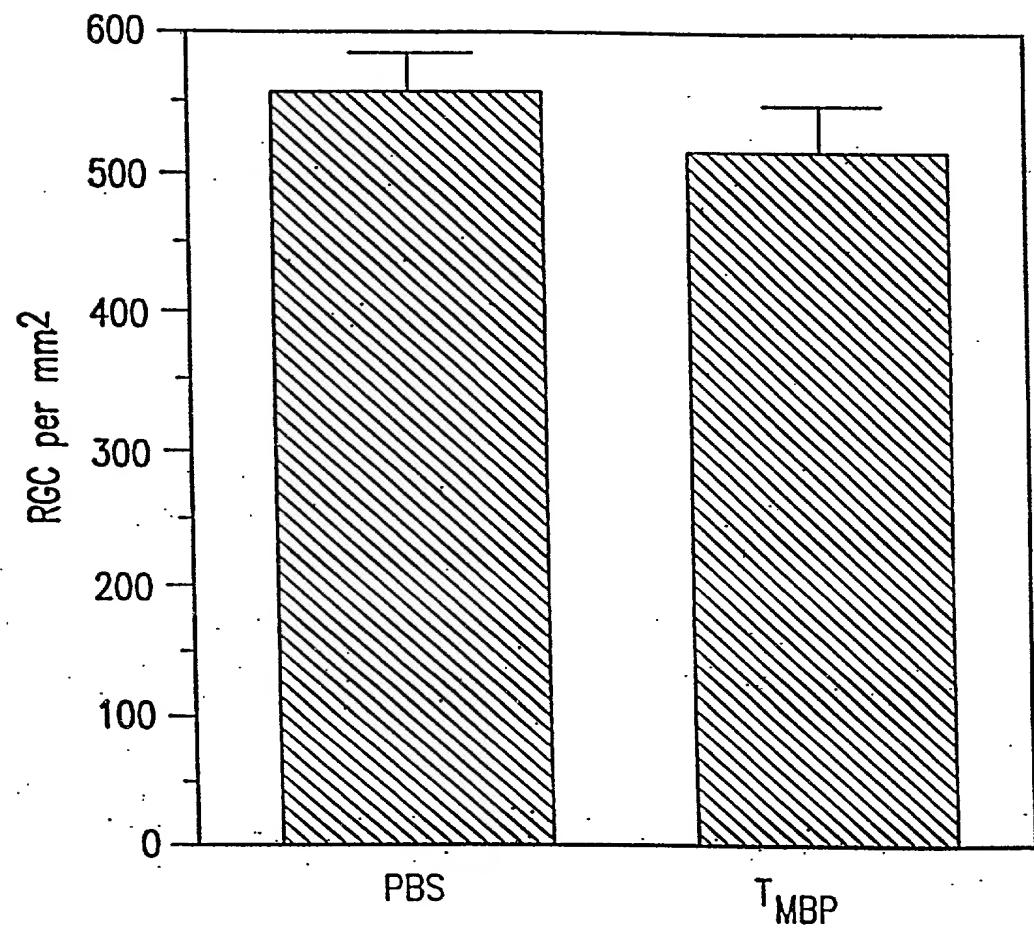


FIG. 4B

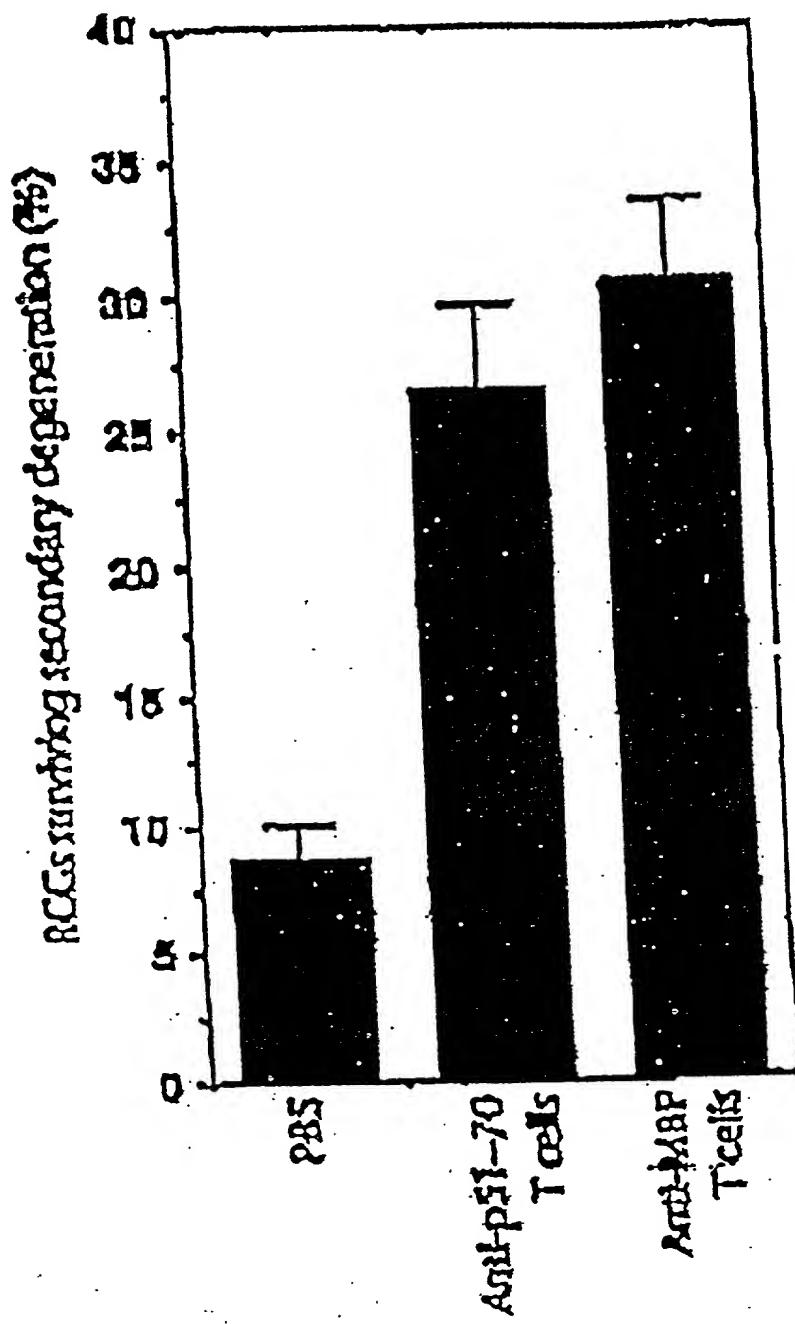


FIG. 5

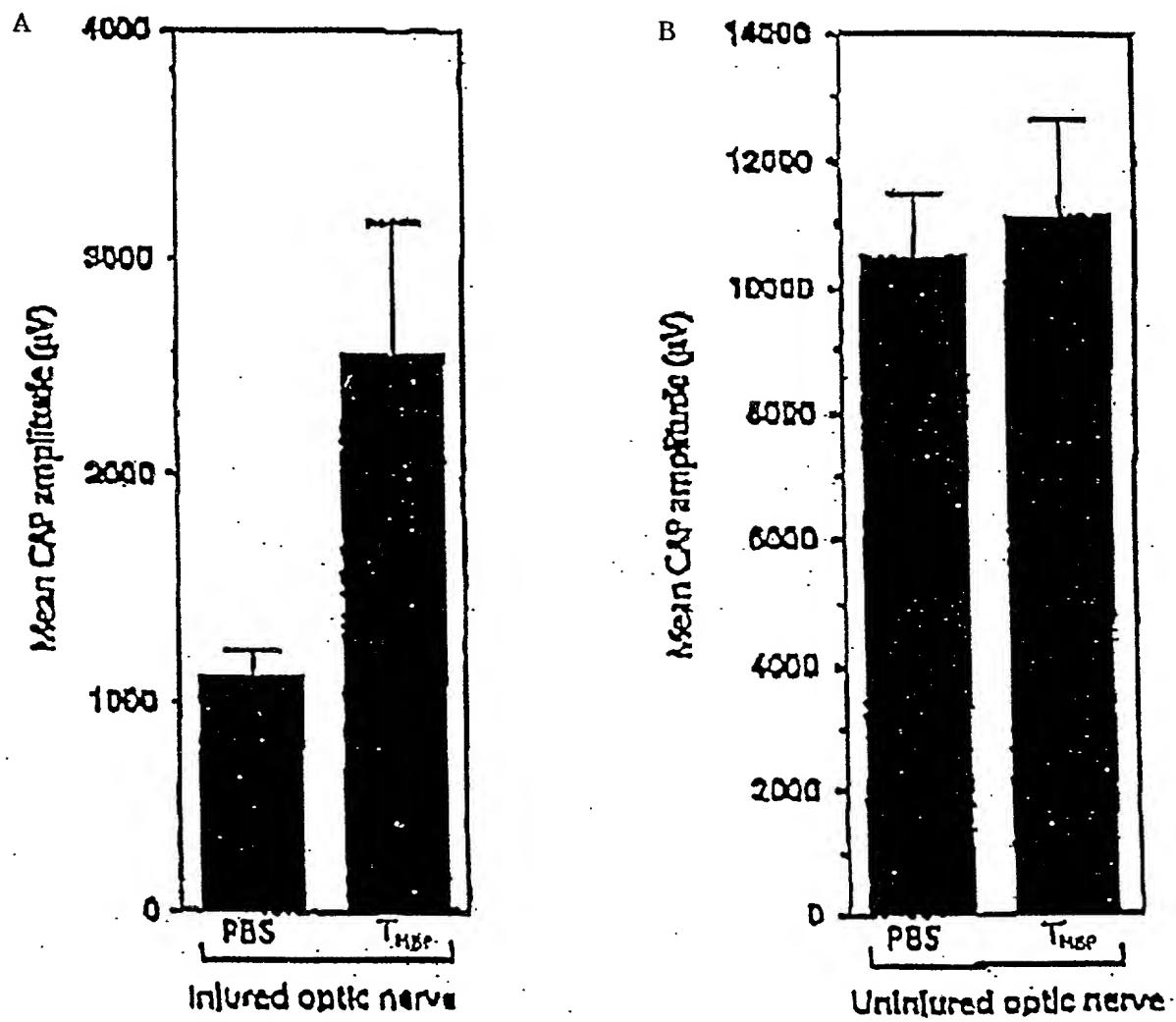


FIG. 6

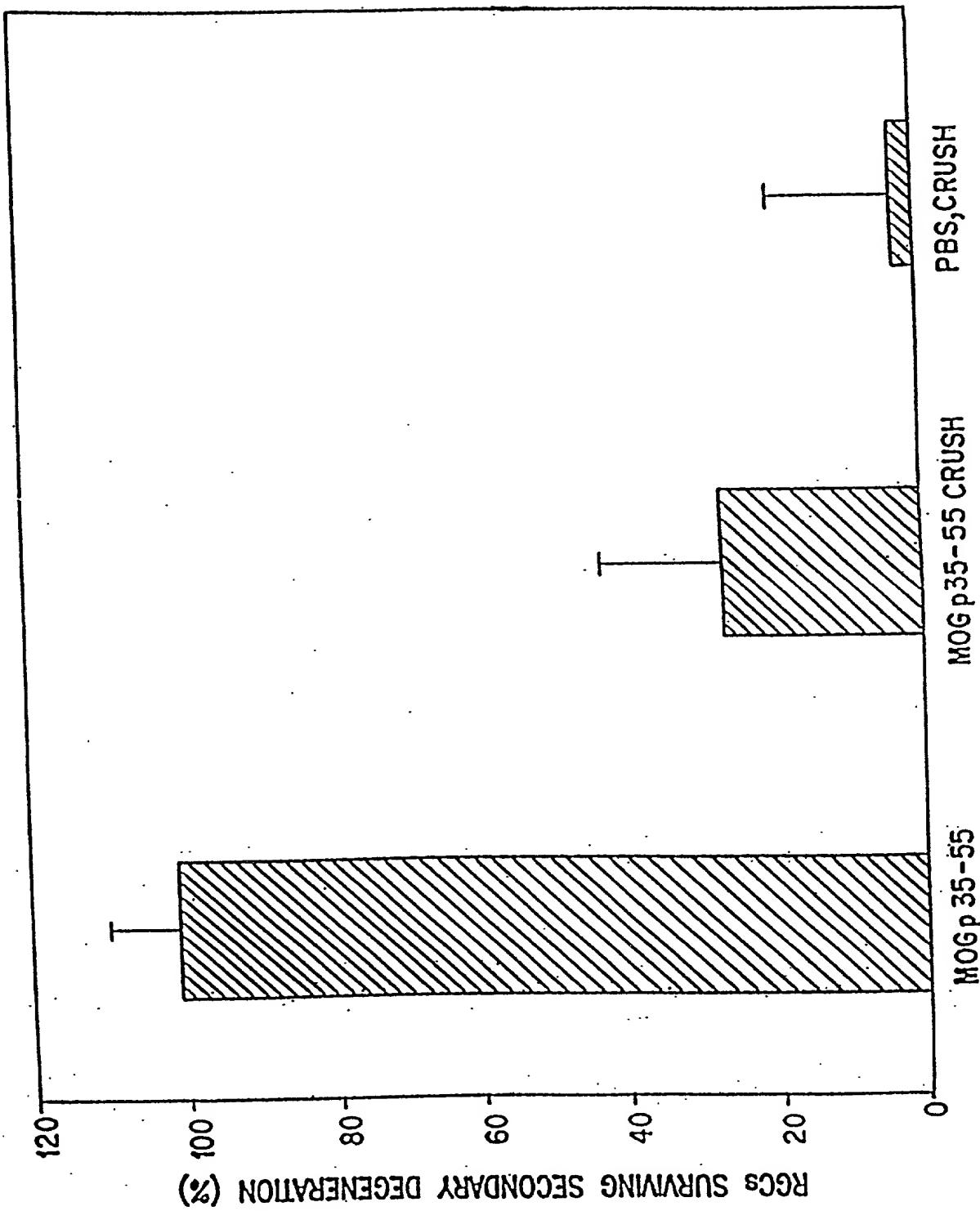
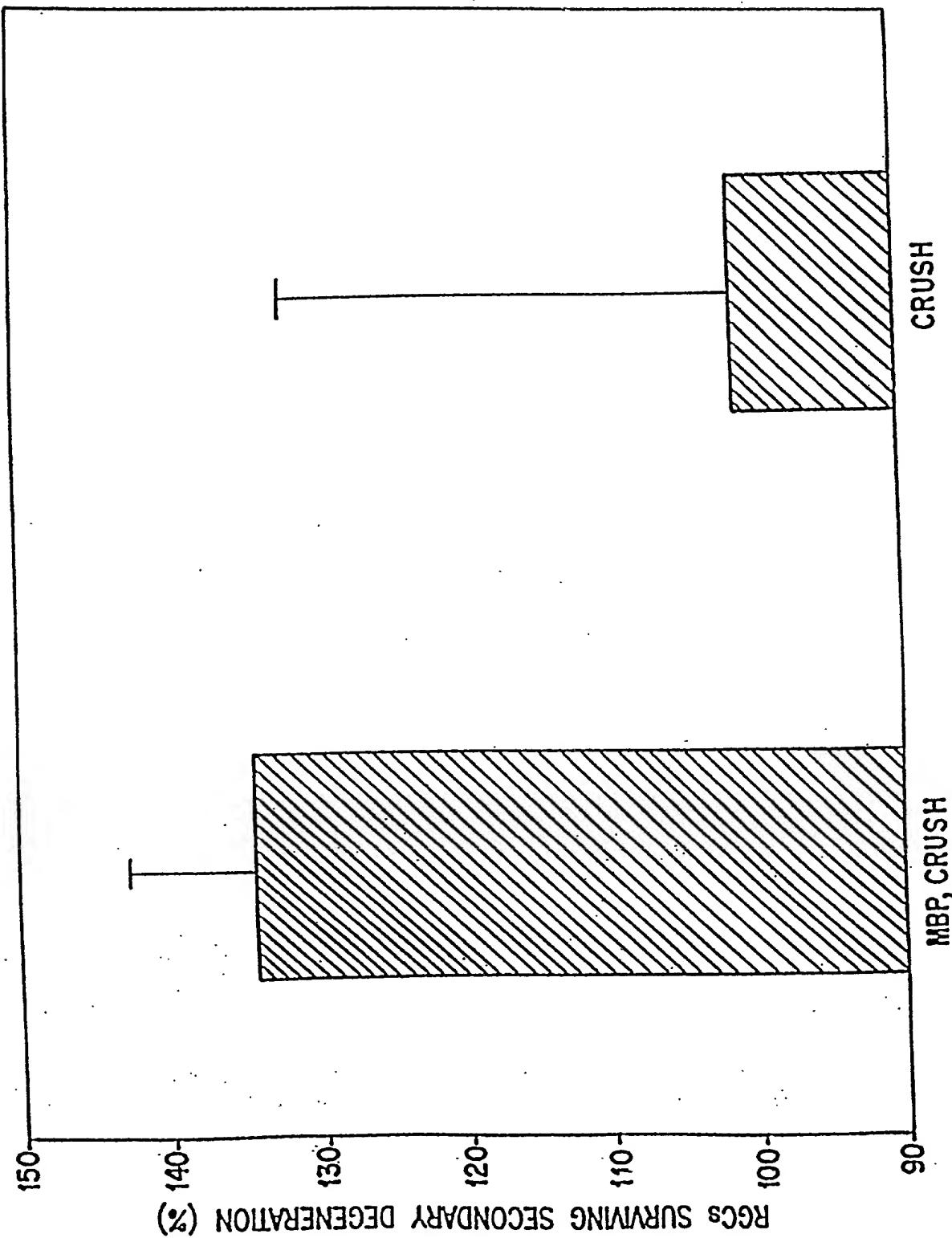


FIG. 7



1 ccaagaagat cccacagcag cttccgaagg cctggatgtg atggcatcac agaagagacc  
61 ctcacagcga cacggatcca agtacttgc cacagcaagt accatggacc atgcccggca  
121 tggcttcctc ccaaggcaca gagacacggg catccttgac tccatcgccc gcttcttag  
181 cggtgacagg ggtgcgccc agcggggctc tggcaaggac tcacacacaa gaactacca  
241 ctacggctcc ctgccccaga agtcgcagag gaccaagat gaaaacccag tagtccactt  
301 cttcaagaac attgtgacac ctcgtacacc ccctccatcc caagggaaagg ggagaggcct  
361 gtccctcagc agatttagct ggggaggaag agacagccgc tctggatctc ccatggcaag  
421 acgctgagag cctccctgct cagccttccc gaatcctgcc ctccggcttct taatataact  
481 gccttaaacg ttttaattcta cttgcaccaa atagcttagtt agagcagacc ctctcttaat  
541 cccgtggggc tgtgaacgcg gcgggcccagc ccacggcacc ctgactggct aaaactgttt  
601 gtcccttttt at

FIG. 9

1 gaaaacagtg cagccacctc cgagagcctg gatgtatgg cgtcacagaa gagaccctcc  
 61 cagaggcacg gatccaagta cctggccaca gcaagatcca tggaccatgc caggcatggc  
 121 ttccctccaa ggcacagaga cacgggcata cttgactcca tcgggcgtt cttggcggt  
 181 gacaggggtg cgccaaagcg gggctctggc aaggactcac accacccggc aagaactgtc  
 241 cactatggct ccctgccccca gaagtacac ggcggaccc aagatgaaaa cccctgtac  
 301 cacttcttca agaacattgt gacgcctcgc acaccacccc cgtcgcaggg aaaggggaga  
 361 ggactgtccc tgagcagatt tagctgggg gccgaaggcc agagaccagg atttggctac  
 421 ggaggcagag cgtccgacta taaatcggt cacaaggat tcaagggagt cgatgcccag  
 481 ggcacgctt cccaaatttt taagctggg ggaagagata gtcgctctgg atcacccatg  
 541 gctagacgct gaaaacccac ctgggtcccg aatccctgtcc tcagcttctt aatataactg  
 601 cctttaaaact ttaatccac ttggccctgt tacctaatta gaggatgaa cccctccct  
 661 aatgcctcg gagggtgcg cgtatgggg tcaggccacg gcaatattcc ggcaatttcc  
 721 ggccaaacagt taaatgagaa catggaaaaca gaaaacgggtt aaaaactgtcc ctttctgtgt  
 781 gaagatcaacg ttccctccccca cgcaatgtgc ccccaacacgc acgtgggtct tcagggggcc  
 841 aggtgcacag acgtccctcc acgttcaccc ctccaccctt ggactttctt ttccgtgg  
 901 ctccgcaccc ttgcgtttt gctggtaact gccatggagg cacacagctg cagagacaga  
 961 gaggacgtgg qccggcagaga ggactgttga catccaagct tcctttgtt tttttctgt  
 1021 tccttctctc acctcctaaa gtagacttca tttttcttaa caggattaga cagtcaagga  
 1081 gtggcttact acatgtggga gctttttgtt atgtgacatg cgggctggc agctgttaga  
 1141 gtccaaacgtg gggcagcaca gagagggggc caccctccca ggccgtgct gcccacacac  
 1201 cccaatttagc tgaattcgcg tggcagag ggaggaaaag gaggcaaaacg tgggctggc  
 1261 aatggcctca catagggaaac agggtcttcc tggagattt ggtatggaga tgcataaggc  
 1321 gtggcctctg gacgtcaccg ttgcctgca tggggcccc agagcaggct ctatgaacaa  
 1381 cctcgtttcc aaaccacacg ccacagccgg agatccagg aagacttgcg cactcagac  
 1441 agaagggttag gacttctca gacgcctcg cagccgcgc agtcgccccat agacactggc  
 1501 tggacccggg cgtgtggca gggcagtgc acgtggcca gcaataacc tccctgagaa  
 1561 gataaccggc tcatttactt cctcccgaaa gacgcgtgtt agcgagttagg cacaggcgt  
 1621 cacctgtctcc cgaattactc accgagacac acgggcttag cagacggccc ctgtatgg  
 1681 gacaaagagc tcttctgacc atatccttca taacacccgc tggcatctcc ttccgtgg  
 1741 ccctccctaa cctactgacc caccttttga ttttagcqca cctgtgattt ataggcttc  
 1801 caaagagttc cacgctggca tcaccctccc cgaggacggg gatgaggagt agtcagcgt  
 1861 atgccaaaac gcgtttctt aatccaaatc taattctgaa tggctgtt gggcttaata  
 1921 ccatgtctat taatataatag cctcgatgtt gagagagttt caaagaacaa aactccagac  
 1981 acaaaccctcc aaattttca gcagaagcac tctgcgtcgc tgagctgagg tcggctctgc  
 2041 gatccatacg tggccgcacc cacacagac gtgtgtgac gatggctgaa cgaaaagtgt  
 2101 acactgttcc tgaatatttga aataaaaacaa taaactttt

FIG. 10

A. 1 taatatctag ggktttgact ctgaccgtg ttggggctct cacttcatgg cttctcacgc  
 61 ttgtgctgca tatcccacac caatttagacc caaggatcg ttgaaagttt ccaggacatc  
 121 ttcattttat ttccaccctc aatccacatt tccagatgtc tctgcagcaa agcggaaattc  
 181 caggcaagcc tttagggaaaa aaggaaaaac aaagaaaaatg aaacaatgg cagtgaaagg  
 241 cagaaagaga agatggagcc cttagagaag ggagtatccc ttagtaggtg gggaaaagg  
 301 gaggagaagg ggaggaggag aggaggagga aagcaggct gtcccttaa gggggtggc  
 361 tgtcaatcag aaagccctt tcattgcagg agaagaggac aaagatactc agagagaaaa  
 421 agtaaaagac cgaagaagga ggctggagag accaggatcc ttccagctga acaaagtca  
 481 ccacaaagca gactagccag ccggctacaa ttggagtcg agtcccaaag acatggtaa  
 541 gtttcaaaaa cttagcatt gaagattca gaggacacag g

B. 1 ctgctttcag agcctgtgac ttcttggtg cctctcctgt ttctcagcaa catggcatag  
 61 ggcctggat accaggtctg gggatctcg ggactcttag cacttaaga cacatgtgtt  
 121 cccaggccct ggtgtgttcc tctagtgcc gaaagatgtt tcatgcttg ctgactttgt  
 181 ataaaagtctg ttttagtgc ttttgacaga atctcagctg ataaatggg gtggggacat  
 241 tagccaaatgc gcatatagg aggacaaaac tgcacataaa atgttccaaa atcattaaagc  
 301 ctgcattttt attattggga gtaataatcaa acctcctatt ttccaattttt catttctgt  
 361 cctgtgctag ctccatcctg tttggactgc tcctccata tggtaactaa gaagaatcaa  
 421 gcattcttg caacaaatac acacgatgtc caaaaatgtc caggagcatc caattccaa  
 481 agtttcctcc acctggaaatg ctcttcatgc taaaatcctg tctgacaataa ccagcatctc  
 541 tggcctgcac tcatccctc ctggaaactcc aagtgcattt accctctgtt accacttaact  
 601 tggctgcctg aattgttagt tgaaaatatt aggtctactt agctaattt tcctcaggaa  
 661 attaaagact cccatatggc agagtctgt tctttctct cttcatatcc cgtataacac  
 721 ccagcataat gctggcata tagtgatgat tccataaataa gttgatgaat gactaaaata  
 781 agcaagcaaa caaacagact agaacaataa gaaagaagg actggatitc ataatctctc  
 841 tggcttgcta tttgaattgc tgaatttta ttattttta aatattttt aaattctggc  
 901 aataaaaggt aaggatttat ttcttttctt tctttttttt ttcttgaga cagagtctcg  
 961 ctcttactgc ccaggctgga gtaaatggc gcaatctgg ctacggcaa cctccgcctc  
 1021 ctctgggtt taacagatc tctctgtcgc gctcttgat tagctggat tacaggcata  
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 1141 ggctggctt gaactcctga cctcatgtga tccacctgc tcacggcaa aaagtgtctgg  
 1201 gattacagggc atgcjccacc gtccccggcc aaagatttat ttcaagaat gaaacaaagt  
 1261 aaggattctg ggtcaatctc acatgtctaa agccaaaacc tctagccgt cctgcttttt  
 1321 gacttcggag tgcccactat ctccgaggct gtgagcacag ggcctggcag aggggttga  
 1381 gtggcatgag ctacctactg gatgtgcctg actgtttccc ttcttcttc cccaggctt  
 1441 ttagagtgtt gtcgaagatg tctggtaggg gcccccttg ttccctgtt ggccactgg  
 1501 ttgtgttct ttgggggtggc actgttctgt ggctgtggac atgaagccct cactggcaca  
 1561 gaaaagctaa ttgagaccta ttctccaaa aactaccaag actatgagta tctcatcaat  
 1621 gtgttaagtac ctgcctccc acacagaccc atctttttt tccctctctc catcctggag  
 1681 atagagaact cttagtacc ttagtaacta gcagggact ggggtggagc cagaccggat  
 1741 tcccgagttct tccctctgtg ca

C. 1 ctagaaaatc cctagccttg ttaagggtct cgctctgggt tatacctcac ttatgtcggg  
 61 aaagaagcca ggtttcaat taataagatt ccctggctc gtttgcctac ctgttaatgc  
 121 aggatccatg ccttcagta tgtcatctat ggaactgcct ctttcttctt cctttatggg  
 181 gccctcctgc tggctgaggg cttctacacc accggcgcag tcaggcagat cttggcgcac  
 241 tacaagacca ccatctgcgg caagggcctg agcgcacgg taacaggggg ccagaagggg  
 301 aggggttcca gaggccaaca tcaagctcat tctttggagc gggtgtgtca ttgtttggga  
 361 aatggctag gacatcccga caaggtgatc atcctcagga ttttgtggca ataacaaggg  
 421 gtggggaaa attggcgcg agtctgtgc ctgcgtccca cccaaggctg ggtctctc  
 481 agggcctgg catttgatg aggaagcgat ggctgcagcc gaacgagaag gtcaaggaaga  
 541 acgtggtgcc cagctggctt agcctcacct ttcaaagggtt ccctaagcaa atttcttc  
 601 aaaacagaaaa gcatgagttt tggggatgc tttgtacaat cagaccatc ctaagccatc  
 661 tgggtgtatc ctttggttcc cttcttagta ggtaccacaa ggtggatct aactggacaa  
 721 gagtctaaaa tgctgctcat gtgattgaga cttgggacc tgagctraga gggaggatgg  
 781 ataataaaaa ttaaataata actccaaggt aaatttacaa tgggtctgg

D. 1 gatcctccctc attcttcccc tacccattcc ccccacccctc cgttatactg gggccaggtta  
 61 tctagtagat actgccaatt acccttgca gaggtgcct gctactaat ttatattttggg  
 121 ggagmccct ggaacctggt ttaatgtct ggcacacgc acttccagga tctccctagtt  
 181 tgtgtttcta catctgcagg ctgatgctga tttctaaacc acccatgtca atcatttttag  
 241 tttgtggca tcacctatgc cctgaccgtt gtgtggctcc tgggtttgc ctgtctgtct  
 301 gtgcctgtgt acatttactt caacacctgg accacactgc agtctattgc ctccccagc  
 361 aagacctctg ccagttatagg cagttctgt gctgatgcca gaatgtatgg tgagtttaggg  
 421 tacgggtgt ttggctctcc taccctactt ggaagacta tatattttgtt tattttctta  
 481 gtgtaaaggag ggtgggtgatt atgagaaaaaa tataatgtat gtaatgattt ggtcttagtt  
 541 tattaaatct tccctactga aaccagagag gtttcttccc ccggaaggaa acttggaaat  
 601 ggtgggagtt ttcttggcca ttccatgg cctactctag ttgactgctg ttacacaaccc  
 661 caaagcagca catttcaata acaaacaacaa ggtttsacca ctgttcaata ccacccctc  
 721 tttttgtaa acctgttagaa aagaggatcc taattgttgg tagmatccaa mtttacagcc  
 781 aggataatta gagatggaaag aaggctctg gggaaagtc tccatgtggc cccgttaactc  
 841 cataaaagctt accctgctt ctttttggat cttacttagg tggatccca tggaaatgtt  
 901 tccctggcaa ggtttgtggc tccaaacctc tggatccatctg caaaacagct gaggtgatgt  
 961 ggttattttgg gttatatttac aaggagatg ctaataccat acaaattaca cccatggcc  
 1021 tcaattttaa ggactgaaag ttcccttgc ctggattttgc aattagccga ttgccttcta  
 1081 caacatgttgc gctaagtgtt cctgagccaa tgagcataga aggtaaaaca cctctttct

E. 1 aatttagcaca cagaaaggat atccaaacaca tacaaggctg tnntcatggc ctacactggc  
 61 gcatattact gctgttgcaa gaaacatttc ttcttcctt tttcattttc ctgcagttcc  
 121 aatgacccctt ccacctgtttt attgctgcattt tggggggc tgcagctaca ctgtttcc  
 181 tggtgatgtt actttgaatg atttggccaa gtaaataggc ctgagatagt tgggttaca  
 241 gctattctga aaggcaagaa ggttagactgc ttccatcctt gaaatgctgg aggaa

FIG. 11F



3061 gcgacagaga gtaagactgt ctaaaaaat aaatgaataa ataaaaagga agaagaagaa  
 3121 gaagaacaat tgcaatcctc cctggctca gaatgtcatt taaaagtca gtgtttctt  
 3181 cttccctgt tttgaagcag cccttctcat gacaggctt cttgccaagg ttccctctga  
 3241 ctttaaatct cttccctttt gttgttggc cagggcattt cagagtataa ggaccaagac  
 3301 accctatccg ggctctggc ggggatggaa tggatggc atgtcgata tctcctggg  
 3361 agaacgctac aggcatggag gtgggggtt accccccccc cttctttagg gtggttcatc  
 3421 tttacagaaaa tggcaaggac caagatggag accaggcacc tgaatatcgg ggcggacag  
 3481 agctgtgaa agatgtt cttgtggggaa aggtgactt caggatccgg aatgtaaagg  
 3541 ttcagatgtt aggaggttt acctgttcc tccgagatca ttcttacca gaggaggcag  
 3601 caatggaaatt gaaatggaa gttgagttt gccatataat attaggtatt aactgttggg  
 3661 tggcaagaaa caattattct ctaacttgcg atgagatccc tcaacccaaa catctcagtc  
 3721 ctggaaatgtt tttccataaa aatgtacaca tcaataaaaca gaaactcatg cttagggatg  
 3781 tctgttgcattt cattatttcg agtagcaagg aaattggggat caaaatcaat gccttgagt  
 3841 aggttaatgtt cagaatgttac aatgttagggc atactgtgaa tattatgcag ggattaaaaa  
 3901 gattattttt gcaactaggcc agatggttt gggggctt ctaaggatattt attgagttat  
 3961 aagagcaagc tgctgttagg tacaaaaaaca aaaacaaaaac cctaggccat ggtgggttgc  
 4021 ctcgcagctt ctcaggaggc tgagacggg ggcggctt agcccgaggg tttcagtt  
 4081 cagtgcgttca tgattgcacc actgcactcc aaccgggtt gacagacaaa gacccctcacc  
 4141 cccactccctt acccgctctt aaaaaaaaaa aaaaacaaaaa caaaaaaaaacc cttggccca  
 4201 ggcgcgtggc tcacgcctgtt aatcccagca ctgtggggagg ccgggttccatcagatcaca  
 4261 ggtcaggaga tcgagaccat cctggctaaa acggtaaac cccgtcttataaaatata  
 4321 aaaaaaaaaaaa aaaaaattttt gccaggcatg ttagcaggcg cttgttagtcc cagactactg  
 4381 ggaggcttagt gcaggagaat ggcgttaccc cgaaagcgga ggttgcgttgg agccaaaatc  
 4441 cttccactgc actccagcat gggggacaca gcgagactcc gttcaaaaaa aaaaaaaaaaa  
 4501 accctgttattt tttgtggcaca cacaacacaca cacaacacaca cttttttttt  
 4561 gtgaaataagc aagttaaatca aatgtctaaa tataattttt gaaaggatgttcc  
 4621 ggctgtactt ccacttattt attctgcaga attgcagaat ttctttttt tttcttttct  
 4681 ttctttttttt ttttttttgg acacagatgc tcgctctgtt acccaggctg gatgtcaatg  
 4741 ggcgcctccg cctctgggt tcaagtgtt ctcctgcctt agcctcccgat gtagctggg  
 4801 ttacagggtgc ccaccaccac acccagctaa tttttgtatt tttagtagag acagggttcc  
 4861 accagggttgc caagggttgc ctcaacttcc tgacctcagg tgatccactc gcctcagact  
 4921 cccaaatgtc tggattaca ggcgttaccc atggtcccg gcctcagaat ttcatttca  
 4981 acatgttttgc catgtgggtt gatttggag aatattttttt gctctatcgc aggtgatata  
 5041 agatgtggac aaggttgc aatgttgc cgttgggggg ggagcttttga aagtacttgc  
 5101 gagttactaa actgttttgc gaggctgggg gtcagatctt ctgccttttcc  
 5161 cctgcagtgc aaacatcaga caattgtatca ctattgtatc ttggagggtgg gatgtaccat  
 5221 tgcagtgcgtt ggaccagaag atggcatgtt atgtggaaaca acaaaggactt atttcttagag  
 5281 actgcctgc tggatgttgc aatagcttta tttgtctcgtt aatgttcttcc atacagctgt  
 5341 ttttattttttt gaaatttttgc ttggccaaaa gtttgcgtt gatgttgcgtt  
 5401 gatgttttgc tttctgcctt aacaacttcc tagtgcgtt actgccttc ccaacaaaact  
 5461 ccctcgtttt caccacacca aaaaaggaaag acaagccggc tgcgggtggc cacacctata  
 5521 atccccaaaatcc tttggggggc cgaggcggtt ggatccaccc gaggctggg gttcgagact  
 5581 agcctgacca acatggggaa accctgttcc tactaaaaac aaaaaatttttgc  
 5641 tggcgatcc ctgttaccc agtggggagg ctgaggcagg acaatcgctt gaaaccccgga  
 5701 ggcggagggtt gcaatgttgc aatgttgcgtt cattacactc cagtttttttgc aagaaaatgt  
 5761 gaactccatc tccaaaaaaa aaaaaaaaaa aacaaggaa acaaaaagaa aagcagctaa  
 5821 agacttttgc tcaaggggaga aatgttgcgtt ttgggttgc atccacatcc  
 5881 ttcccaccc ttcgttgc tgcctaaatcc actgttttac aagtaataaa gggacgctt  
 5941 gtctaggctt tggagccagg aatgttgcgtt aatgttgcgtt atgagatgaa gtaatggat  
 6001 tatttgcgtt ctcagggttgc actacacttcc ctcttcttca gaaaggatcc taatttctt  
 6061 tttttttttt gtcatttttgc gatgttgcgtt actgttgcgtt tcaatcc

FIG. 12 (cont.)

6121 tctttttaatt cttcattatg aaacataaaaa acaaatgcca ggcgccggag ctacgcgcct  
6181 taatcccagc actttggag gccgaagccg gcagatcacc cgggtcagga gttcgagacc  
6241 agcctgatca acatggagaa accccgtctc tactaaaaaa tacaaaatta gctaggcgty  
6301 gtggcacatg ccagtaatcc cagctacttg agagactgag gcaggagaat cgcttgaacc  
6361 gggaggcaga ggttgcggg agccaaagatc ggcattgc actccaggct gggcaacaag  
6421 agcaaaactc tgtctcaaaa aaaaaaaaaacc acatacaac cagagataat attataatga  
6481 gcctccaagt gcctaccacc ttgctgcagc acttgtcaat ccagggacca cccacctcac  
6541 cggctccccca ctcatattca ccctccccca ctcaattact gaggttaaatc ctaggcagca  
6601 tgcatttttc tttttttct ttttattat tttgagacag gatctgtctc tgtcacccag  
6661 gctggagtgt agtggcatat ctctgctcac tgacgcctc gcctccccc gagaagccat  
6721 cctccacact cagccatcat agtagctggg accacaggca cacaccacca cacactgcta  
6781 atgtttttaga tttttttag agactgggtt ttacatgtt gatcaggctg gtctaaact  
6841 cctaggctca agcaatccctc ccacccctc ctcggcctt gctagaatta caggcgcag  
6901 ccactgcacc cagcgaagaa cacttttaa aaaataaata ggcggggcgc ggtggctcac  
6961 acctgtatc ccagtagttt gggagccaa ggaggcgaa tcataggtc aagagattga  
7021 gaccatctta agtaacatgg tggaaacccca tttctactac aaatacaaaa acaaattaa  
7081 cctggcgtgg tggcaggcgc ctgtagtccc agctacttgg gagctgaggc aggagaatgg  
7141 agtgaacccg ggaggcggag cttgcagtg a gctgagatca tgccactgca ctccccctg  
7201 gggcaacaga gtgagactcc caaaaaaaaaa aaaaaaaaaa cccccctccc acacacaata  
7261 atataaataaa ataaataacc acaatactat tatcacatct tacaactca acaaaaattt  
7321 cttaatatca tcaaataccca agtttgggtt caaattttcc tgattgttc ataaatatac  
7381 tcttacagtt ggtttctttt agcgagatc aaatgagacc cacctgttga ccttgcct  
7441 tagggtttcc cagggctctga attttggta cgacattccc atgttgcata gtaatacggt  
7501 cctccatgcc ctgtgtttt ctgtaaactg atagatgtgg aggtgcaatg acatttgg  
7561 ttgatttact ttggcaaaaata tagttcatca gtgatactct atacttctg ttgcttaca  
7621 tccggaggct gataatgtct gctttctct ctttctaat tatttggtaa aggaaaaatg  
7681 tgggggggtt ggagaaaaaaa acccttaagt acataactcgc taaatcacat tgctacaggt  
7741 aacttccatt aagaacttga aagtaaagt agctgcatt tcccctaggg aacacaaatga  
7801 tagacaggag ccttagtctc cagcttgaag gattgttaatt atacctaagc aaccctctg  
7861 gaccagttt atgttattag ctgtatgtt cccctacatt tgatcatttctt atcttactt  
7921 agctccctta aagcagagat caagatggaa agggttcag ctgcagcatg gcacatggag  
7981 attagagtgg ggcttttggc tgctgaggag cagacctaga atggaaata gatggaggg  
8041 acagaagtga aggtccccct ccctcatttc tcaacactact ccacatctcc aggtctgcac  
8101 atctgttcaag ttactgaatc ctgtgttaaagc tacottctt ttcttttttcc ttttatttt  
8161 ttatttattt ttttttttag atggagttt gctttgtta cccaggctgg agtgaatgg  
8221 tgcaatctcg gtcactgca ccctccaact cccagggtca tgcaattctc ctccctcagc  
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8401 ccacccaccc tggccctccca aaatgctggg attacagggt tgagccacca tgcccgtgt  
8461 aaactaccc ttttttttttctt ttttttttttctt ttttttttttctt ttttttttttctt  
8521 cttccgcaag ctgtatgaaat ttttttttttctt ttttttttttctt ttttttttttctt  
8581 agtgtctcac tctgtcaccc aggattgggtt gcagtggcac gatcatgggt cattgcagcc  
8641 tccacccccc aggtcaagt gatcccttcg actcaggctc ttgaatgtt gatcatgggt  
8701 gcttggtca ccatggccag ttttttttttctt ttttttttttctt ttttttttttctt  
8761 ttttttgcctt agtctggccct ttttttttttctt ttttttttttctt ttttttttttctt  
8821 cccaaatgg tgggattaca ttttttttttctt ttttttttttctt ttttttttttctt  
8881 acacccaaata aaacattttat ttttttttttctt ttttttttttctt ttttttttttctt  
8941 agggtgaccc tatttttttttctt ttttttttttctt ttttttttttctt ttttttttttctt  
9001 gatttctgtt acttttctaa ttttttttttctt ttttttttttctt ttttttttttctt  
9061 ttttttggaa tggagtttcc ttttttttttctt ttttttttttctt ttttttttttctt  
9121 ctcacccctt cctccgcctc ttttttttttctt ttttttttttctt ttttttttttctt

FIG. 12 (cont.)

9181 ctgggattac aggactgtgc caccacgtcc agctaatttt gtatttttag tagaaacagg  
 9241 gtttctccat gttggtcagg ctggtctga actcccaacc tcagggcgtc cgccgcctc  
 9301 ggcctccaa agtgcgtggg ttacaggtgt gagccaccgc acctggccaa tatttgcgt  
 9361 ttttattgac gacaaagtca aaggttctct tcatttattt gtggtgtatc gcctacaagc  
 9421 ataattaaaa taaacactaa atttcagtt aaagttact gaaaataaat atgtatttt  
 9481 tattccctat ttaagcttg aatccctga cttcctatac cattaccact gtcctagttc  
 9541 aggttcatgt tgggtttac ttaattgtt atcacagtct ctaacattt ctcctatgt  
 9601 tctccagtcc tggtaggtgt aatctgacg tggtcacttc tcagcttggatccttgcgt  
 9661 gcaccaccac agccttgaac tacatattt aatatacatat ttattttcg taaactttaa  
 9721 actgaaattt agtggtttatt ttaattatgc ttgttaggcga tacaccacaa taatatgaag  
 9781 agaaccttgcg acttgcgtt caataaaaaag tccctgggg ggacttcaga tgtaagtc  
 9841 ttagctgtc gttaaaactc cccagggtg acccaataca caatcttgac tttaaaccac  
 9901 ttgtcattct aaatcactag catttcctgg aaaaaaaaaagc catttttct tcagggctaa  
 9961 gtcaggagac caattctgtg tcaccttctt tgaatcctga tgatattcac ttcttattt  
 10021 gacctgatatt attggggccc agacaccatg ctgagtttgc gggattcagc tctggacaat  
 10081 gtcaaatgtc agtccctgcct ttcagatct ttctacttgg tgagccctgg agtgcgtgg  
 10141 ctccctcgcc tgctgcctgt gctccctctg cagatcactc ttggcctcg tttccctgc  
 10201 ctgcagtgaca gactgagagg tacagggcag agggtgggtg gatcaggatc cttctttaa  
 10261 atgagctggc ttcttggagc tacaccactt aacatgtatt tggtagtgac ttctgggtt  
 10321 agaagttctt ctcactattt agtgataaaag aaaaaaaaaata actccatgtat gaaagagtt  
 10381 tacatcttac ggaatgtctt catatgaata atccgaccta gcattttccat atgagctaac  
 10441 tatgccatata agtaacccca ttgttacagag gatacaactg aggccaggag tagttcagtg  
 10501 acttactcaa accgatataa ttatataatgt gtagagctga ggcctctgtc tcatacctag  
 10561 cagctccatg caacttggga gagggtgagc ttccaaatgtca gacagggtcta ggcttattag  
 10621 agtttgaat aaagataactg aagtggaaatg ctctaccaca cagtaggggt tcgaaaatttgc  
 10681 ttccctttt ctccattcaa cacttggagc tcagggttgc ctgtgtatgc agtccctt  
 10741 ttgtccctag agtttctt cttttcccttacc aagtgttcc ccaatgccag  
 10801 agcaggaaga gtcttcactc ctcccaatgc cccacccccc atttggttact aagaggagag  
 10861 gagaaatgtt caaggagggt atggggatg ttctgggggatg atgggtgttgc tgccatcaa  
 10921 caacaaatgtc ctttctctca ctttgaattt atcccagatg cttgtttgtt tacttcttcc  
 10981 acacaaaaaa aggcttca ggcctatggc tgagcagaaa gaatctgaat tttagagtca  
 11041 ggcagcttgg gttgaattt catctcaggactt actgaactct atagcaaaat tcttagattt  
 11101 tccaaatgtc agtttgcctt tctgtcaat agagaaaaca tccttcgtcc taaattgttag  
 11161 ggaggatata agtcatgca agtgcctact acaaatccag tcacaaatgtact  
 11221 cactaaatgt tcagcttctc ctttgcattt ctttgcattt ctttgcattt ctttgcattt  
 11281 tggcaacgca gtgggcttggc gcaatgttgc gaaatggaaa tccaaagaaaaa gggcgaaga  
 11341 gcaatgttggc tggtttttttt gcttgcgtt gcttgcgtt ttgttccat tcttttatttgc  
 11401 ctattgtatc tagactatag cttagaaaaag agccgcaccat tttttttttt aatccatgtc  
 11461 tttttttttt ctccatgtt gttttccagg ctggcagaaa atagccttgc caagggggccc  
 11521 aggcttggg tggggggggg tttttttttt ggggggggggggggggggggggggggggggggggg  
 11581 cttttttttt tttttttttt tttttttttt tttttttttt tttttttttt tttttttttt  
 11641 ccaattttttt cttttttttt tttttttttt tttttttttt tttttttttt tttttttttt  
 11701 tccctcttca tttttttttt tttttttttt tttttttttt tttttttttt tttttttttt  
 11761 cttagaaatgt tttttttttt tttttttttt tttttttttt tttttttttt tttttttttt  
 11821 gtggccctact aaatccattt ccattttttt ctcccttgc gaaatccac cggactttttt  
 11881 gtaagttccg gcatgttgc gttttttttt tttttttttt tttttttttt tttttttttt  
 11941 cacctgggggg aacaaggacc ctttgcgtt gttttttttt tttttttttt tttttttttt  
 12001 ttctttttttt tttttttttt tttttttttt tttttttttt tttttttttt tttttttttt  
 12061 gtactgtttt tatgtgcacat gcaatgttgc gttttttttt tttttttttt tttttttttt  
 12121 gttttttttt tttttttttt tttttttttt tttttttttt tttttttttt tttttttttt  
 12181 tcgttttacac gtaaaatgtca tttttttttt tttttttttt tttttttttt tttttttttt

FIG. 12 (cont.)

12241 actgtatcac ctgtacttat atttctgctt tacaaactca ggatgttcc atgagtag  
 12301 aacatgacta atcagagaag acctcataga ggaatagaaa agccaccaag cccactagg  
 12361 aattgacccc tcaaggacat gtttctagc cttttgttc actgcagatt gccaaatgcc  
 12421 taaagataat ggcaacagaa gagcacccaa atatttgtt gataaatgtt gcagacacta  
 12481 gaagggtgtca ttagggcaca gatggtaacct tctctgagca aacttccttc acagctcctc  
 12541 ctcccgaggg tgttaggtgac tctactcttg tcacctggca cacagagttc tatcgatcg  
 12601 ttttagaaat tagaccagtg tgggaccac acacacacac atctttacac accaaagag  
 12661 gaggaatagt atctttgtt tggaggactt gactatgaaa ggtcttaact ccttttgta  
 12721 ccatgaatct ctctggcact ccagtgaagt ctaaaggacc cctttgcaga atgttttaa  
 12781 atatacacat aaaatagaac acataggatt gcaaaaacaa tcattgtact aaaatacagt  
 12841 tatcaacccga taatcacatt tctgtatag taacataaat gtttctttt ttttttttg  
 12901 gaggcagagt ttggcttgc tcacccaggc tggagtgca tggcgcgatc taggctcact  
 12961 gaaaccttg cctccgggt tcaagcgatt ctcagccttc tgtagtagctg ggattacagg  
 13021 tgccgcac cacacccagc taattttgt attttttagta gagacttagt ttacccagg  
 13081 tggccaggct ggcctcgaa tcctgaccc aggtgatcca cctgccttg cctccaaag  
 13141 tgctgggatt acgggcatga gccaccgtgc cccgccccata atattttttt agccaaagta  
 13201 atacattaag taatgttaga gcaagtctaa taacctgtaa ttctttttt tcttccttc  
 13261 ttctttttt ttggatgtg agttttttt agatggagtg caatggcaca atctcgctc  
 13321 actgcaaccc ccaccccttg ggttcaagcg attctcctgc ctcagcctcc caagttgctg  
 13381 gaactacagg cgcatgccac catgccccagc taatttttt attttttagta gagacgggg  
 13441 ttccaccatgt tggccaggct ggtcttgaac ccctgaccc aggtgatctg cctgccttg  
 13501 ccttccaaag tgctgggatt acaggcatga gccaccaggc ccagcccaat aaccttaat  
 13561 ttcaacatac taataaacat aaacagtatt tcaagatttgc tgcaataact ctaatggaa  
 13621 tggaaacatc tggcttgc atttttttt aagtccatgg tactgctcat attgtggta  
 13681 gttgtaaaat gttttgggtt gttttttt ttccaagact tgggggaatg ggtgttgggt  
 13741 ggatcaacaa gagtcttgct ctgtggccca ggctggagtg cagggggcagg atcttggctc  
 13801 actgcaaccc ccgcctccca ggttcaagcg attctcctgc ctcagcctcc tgtagtagctg  
 13861 gcattacagg catgtgccac cacgccccagc taatttttt attttttagta gagatgggg  
 13921 ttccaccatgt tggcttgc ggtcttgaac tcttgcctc atgtccacc cgtctcgac  
 13981 tcccaaggatg tgggattac aggcatgac caccacccat ggcagttttt acatttttaa  
 14041 tggaaagaaaa tggtaatcc agttatttggaa aataaggagg cagttttt ctcatccaa  
 14101 ttcatggact ttctgaattt tggcccccaga gtccttttgt gttcttaggac cccaggttaa  
 14161 ggaacccaaa aagacagggt ggtggggcat gagggggaac acatgttaat ccctgtttgt  
 14221 tctgggtgaac aattcagatc cccacttttct gagggtgc ctcgttggaa taaccctgtt  
 14281 tggtaatttggcccggttgc gacccttttgt tgcttgc tctgttgc atctgctaca actggctaca  
 14341 tcgaagacta gcagggtcag tggctggca gcaggcaaga ccaccaaata gtgggggacc  
 14401 aagtccatgt tggatggaa gccaaaagag aatagaacca ggactcaaga ttaggggagc  
 14461 tgggatttcc ttattcctct gtccttgc ccaacccat gctttctga gaaactgtga  
 14521 agagaaccac ttactggatc tggggatcc cccactggaa agggcagttt gggctactcc  
 14581 aaatgtccat agggaggatg tggggaaaggctt gcttccatcatc ttccactaat cacatattt  
 14641 ttctttttt ttttcaggcc aattccttgc agagctacgt aagttctt ctctctgtt  
 14701 taaggcagaga ataaaaagcc agggaaaggga gacagaagca acaagaggaa gagggggct  
 14761 attggggat cacatttccca gaggaaaggga ggagctggag agcctgggtt gaggaaagac  
 14821 tcctccttggg aggttagaggg caaagaagcc agctgttaga gacacatttgc cagttggcag  
 14881 agaagcttggaa ggcacttccat tctgcccactt gatccatttgc tccttcaactt cccctaaagca  
 14941 ggaatccaa ccttagtggt ctcatgtcc attccacccat gactgcccag tgcttcaccc  
 15001 ctcagatcaa ccattggatc aggaatggag acaagatgac cccaaaggctt tttttcttcc  
 15061 ctagttcaat ggttttatgt tacaactac tgacatactg tttcaagttt atttttctt  
 15121 tttcttagga aatcccttgc ggtgtatgtc acatcttggc aggggtggag gagagcttgg  
 15181 ttggccagggtt atttgcctt ggggacatctt catccatcaa gttgcacact cactggcatc  
 15241 ttgtctatgg ggacattcca atttgcatttgc tcaggaacac tctgaattcc aagtagattt

FIG. 12 (cont.)

15301 gatttccctt cttctgtcat ctacctttc tcttcatttt cccatttttta ttacccttct  
 15361 ttccatattct ctctccagtc ttccacctgg aagccctctc tggctaagga caggcagggtg  
 15421 cccctctctc catcagagga cacctgtact ggagagcaac acaggatggt ctctgccatg  
 15481 aactggaggc caggaatctc ctcaactgaaa attacagtat ggttaactttg caaatggtg  
 15541 ttgtttcttc caagactcca gcccgtattt cgccaaaactg aaaggcatgt gaagggaaagg  
 15601 aagaggaaga gtgcaaaaaca ttgaagagag agctgagtga gctgaagagt gaggatatga  
 15661 gtagcccaa cccaaacctg gagatgggg aaaacctaca gaataactagc cagagctcct  
 15721 ccttgccttg gcagcctact agggacctgg ggaagcaaaa acgaaagctg ggcaacatgc  
 15781 ctgctttaga atgtttctt tctacttaca catcttccac aggtctcaga atctttcctt  
 15841 cctctcatcc ttttctccta tctacatatc tattcagatgta tccactgtttt attcaacaac  
 15901 tactacttga tggtcagaca caaacaaca agcttaggtgc taattaataa agatacgaat  
 15961 tttggccggg tgcgggtggct cacgcctgta atccccagcac tttggggaggc cgaggcgggc  
 16021 gaatcacgag gtcaggagtt caagaccagc ctggccaaca tggtaaaacc ccatctctac  
 16081 taaaataaca aacaattaac tgagcatagt ggtgggacc tataatacca gctactccgg  
 16141 aggctgaggc aggagaatcg cttgaaccca ggaggcagag gttgcagtga gctgagatcg  
 16201 cggccactgca ctctagccgg agtgcacagag taagactctg tctcaaaaat aaataaataa  
 16261 ataaataaaat aaataaaataa ataaataaaaa aataaataata caagtttca taagcacact  
 16321 tctaaccctt tgcgttttat gtatttctt ctttatccac gcacctgtct ccctctactc  
 16381 cagcctcattt accccagagg tcagtcctca gggaaactaa acacaaagaa agagctcagt  
 16441 cagaaaaggcc atttattttt gtttcaagat gtcactgccc tcctttgttt tgcgtttttt  
 16501 gcaggccctc tcttttaggc ctcttctcctt ggggtatgg atcctggggg gagattgatc  
 16561 acctccatgc ttccattctt ccccgccat agtggggaca tcatgagaga agccaagcca  
 16621 ctggcccaagg atcacccggc atttatggg gtcgtctgg cacagtcct tgcctttata  
 16681 gcccctccag tgatccataa gcccctttt ctccccaaag gagaggtcac agatagggca  
 16741 aaggtagctc ttctgcttcc agtgggtctg ctgggtctg accagcctgg aaaatgagct  
 16801 gaaagacttg ctgcaatggg agcagtagtt gggcggctct gtgagggtgc cttctgggt  
 16861 tctggagaga tagatttttct tgctaaaatgta caaaaacaa tggggcaac agaagacatt  
 16921 gagtctttag ggcctcactg gatgagagtt ggtatctggca tcctgacaga gggttccagt  
 16981 gatgggtgcc tgggtctgg tcacagtgatc ttgggttctta agtacagatg cctgggtctg  
 17041 ggccttagga ccctcagttc taaatatggg ttctgggac ctggccactg gtgcatgggt  
 17101 cacatccaaa agccctgtgg tggacctctg gtttctggcg atgggtgtct ggaattcagc  
 17161 ctgggtgcct ggaatccctca aagtacactc ctgggttccca tccactggct cctgggtttt  
 17221 gtgtatcttc tgggtggcgtt tgagctcaga ctgggtccgg aagctcttcc cacacacaga  
 17281 gcatgaatgg ggcggtaac ccagatggac gcccgggtga cgacttagtc cagaagcatc  
 17341 acagtaggtc ttgtcacaga gctgcaaca gaagggcctc tccccaagat gcatgcgtct  
 17401 gtgatagctg aggacttgg ggctccggaa caacttccca cactgactgc agctgttagt  
 17461 cagttggga ttgtgaacaa actgggtggct atagaggtag gagcgcctgc taaaacattt  
 17521 ggcacaggtg tagaaaaa

FIG. 12 (cont.)

1 tttgtatgtc attgcaggat tcatgcttgc cagtgtgtca tctatgaaac tgcctcttgc  
61 ttcttcctt atggggccct cctgctgct gagggcttct acaccacccgg cgctgtcagg  
121 cagatcttg gcgactacaa gaccaccatc tgccgcaagg gcctgagcgc aacgtaaca  
181 gggggccaga aggggggggg ttacagaggc caacatcaag ctcattttt ggagcgggtg  
241 tgtcattgtt tggaaaaatg gctaggacat cccgacaagg tgcattttt caggattttg  
301 tggcaataac aagggtggg gggacaa

FIG. 13

1 ctgtatcagt gctcctcgtc gcctcactgt acttcacgga agagacttgg ttgactggcc  
 61 acttggagcg gaatcaggag acattcccaa ctcagagaga ctgagcccta gtcgccccac  
 121 ttgctggaca agatgatatt ctttaccacc ctgcctctgt tttggataat gattcagct  
 181 tctcgagggg ggcactgggg tgcctggatg ccctcgcca tctcagccctt cgagggcacg  
 241 tgtgtctcca tcccctgccc tttcgacttc ccggatgagc tcagaccggc tgggtacat  
 301 ggcgtcttgtt atttcaacag tccctacccc aagaactacc cgccagtggt cttcaagtcc  
 361 cgcacacaag tggtccacga gagcttccag ggccgttagcc gcctgtggg agacctggc  
 421 ctacgaaact gcaccctgct tctcagcactg ctgagccctg agctgggagg gaaatactat  
 481 ttccgaggtt acctggcgcc ctacaaccag tacacccctt cggagcacag cgtcctggac  
 541 atcatcaaca cccccaacat cgtggtgccc ccagaagtgg tggcaggaac ggaagttagag  
 601 gtcagctgca tggtgccgga caactgccc gagctgccc ctgagctgag ctggctggc  
 661 cacgaggggc taggggagcc cactgttctg ggtcggctgc gggaggatga aggcacctgg  
 721 gtgcagggtt cactgttaca cttcgtgcct actagagagg ccaacggcca cctgtctggc  
 781 tgtcaggctg ccttccccaa caccacccctt cagttcgagg gttacgccc tctggacgtc  
 841 aagtacccccc cggtgattgt ggagatgaat tcctctgtgg aggccattga ggctccac  
 901 gtcagctgca tctgtggggc tgacagcaac cggccaccgc tgctgactt gatgcgggat  
 961 gggatgggtt tgagggagggc agttgctgag agcctgttacc tggatctgga ggaggtgacc  
 1021 ccagcagagg acggcatcta tgcttgctg gcagagaatg cctatggcca ggacaaccgc  
 1081 acggtgagc tgagcgtat gtatgcaccc tggaaagccca cagtgaatgg gacgggtggtg  
 1141 gcggttagagg gggagacagt ctccatccctg tggccacac agagcaaccc ggaccctatt  
 1201 ctcaccatct tcaaggagaa gcagatccctg gccacggctca tctatgagag tcagctgcag  
 1261 ctggaaactcc ctgcagtgc ccccgaggac gatggggagt actgggtgtt agctgagaac  
 1321 cagtatgcc agagagccac cgccttcaac ctgtctgtgg agtttgcctt cataatcctt  
 1381 ctggaatcgc actgtgcagc ggccagagac accgtgcagt gcctgtgtt ggtaaaatcc  
 1441 aacccggaaac cttccgtggc ctttgagctg cttcccgca acgtgactgt gaacgagaca  
 1501 gagagggagt ttgttactc agagcgcagc ggccctctgc tcaccagcat cctcacgctc  
 1561 cggggtcagg cccaaagcccc acccccgctc atttgtaccc ccaggaaccc ctacggcacc  
 1621 cagagcctcg agtgccttt ccagggagca caccgactga tggggccaa aatcgcccc  
 1681 gtgggtgtt tggtcgctt tgccatccctg attggcattt tctgtacat caccaggaca  
 1741 agaagaaaaa agaacgtcac agagagcccc agtttcttag cggagacaa coctcatgtc  
 1801 ctgtacagcc cccaaattccg aatctcttgc gcacctgata agttagagag tgagaagcgc  
 1861 ctgggggtcc agaggaggct gctggccctt aggggggaaac ccccagaact ggacctcagt  
 1921 tattccact cagacctggg gaaacgaccc accaaggaca gctacaccct gacagaggag  
 1981 ctggctgagt acgcagaaat ccgagtcaag tga

FIG. 14

1 masqkrpsqr hgskylatas tmdharhgfl prhrdtgild sigrffggdr gapkrgsgkd  
61 shhpartahy gslpqkshgr tqdenpvvhf fknivtpntp ppsqgkrgl slsrfswgae  
121 gqrpgfgygg rasdyksahk gfkgvdaqgt lskifklggr dsrsgspmar r

FIG. 15

1 mglleccarc lvgapfaslv atglcffgva lfcgcgheal tgtekliety fsknyqdyey  
61 linvihafqy viygtasfff lygalllaeg fytgavrqi fgdyktticg kglsatvtgg  
121 qkgrgsrqh qahslervh clgkwlghpd kityaltvww llvfacsavp vyiyfntwtt  
181 cqsiafpskt sasigslcad armygvlpwn afgkvcgsn llsicktaef qmtfhlfiaa  
241 fvgaaatlvs lltfmiaaty nfavlklnmgr gtkf

FIG. 16

1 maslsrpslp sclcsfllll llqvsssyag qfrvigprhp iralvgdeve lpcrispgkn  
61 atgmevgwyr ppfsrvvhly rngkdqdgdq apeyrgrtel lkdaigegkv tlirirnvrs  
121 deggftcffr dhsyqeeaam elkvedpfyw vspgvlvlla vlpvllqit lglvflclqy  
181 rlrkgklraei enlhrtfdph flrvpcwkit lfvivpvlgp lvaliicynw lhrrlagqfl  
241 eelrnnpf

FIG. 17

**DECLARATION  
AND POWER OF ATTORNEY**

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below at 201 et seq. underneath my name.

I believe I am the original, first and sole inventor if only one name is listed at 201 below, or an original, first and joint inventor if plural names are listed at 201 et seq. below, of the subject matter which is claimed and for which a patent is sought on the invention entitled

**ACTIVATED T-CELLS, NERVOUS SYSTEM-SPECIFIC ANTIGENS AND THEIR USES**

and for which a patent application:

is attached hereto *(if applicable)*  
 was filed in the United States on \_\_\_\_\_ as Application No. \_\_\_\_\_ with Preliminary Amendment filed on \_\_\_\_\_ *(if applicable)*  
 was filed as PCT international Application No. \_\_\_\_\_ on \_\_\_\_\_ and was amended under PCT Article 19 on \_\_\_\_\_ *(if applicable)*

I hereby state that I have reviewed and understand the contents of the above identified application, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information known to me to be material to patentability as defined in Title 37, Code of Federal Regulations, §1.56.

I hereby claim foreign priority benefits under Title 35, United States Code, §119(a)-(d) of any foreign application(s) for patent or inventor's certificate listed below and have also identified below any foreign application for patent or inventor's certificate having a filing date before that of the application on which priority is claimed:

EARLIEST FOREIGN APPLICATION(S), IF ANY, FILED PRIOR TO THE FILING DATE OF THE APPLICATION			
APPLICATION NUMBER	COUNTRY	DATE OF FILING (day, month, year)	PRIORITY CLAIMED
IL 124550	Israel	May 19, 1998	YES <input checked="" type="checkbox"/> NO <input type="checkbox"/>
			YES <input type="checkbox"/> NO <input type="checkbox"/>

I hereby claim the benefit under Title 35, United States Code, §119(e) of any United States provisional application(s) listed below.

APPLICATION NUMBER	FILING DATE

I hereby claim the benefit under Title 35, United States Code, §120 of any United States application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of Title 35, United States Code §112, I acknowledge the duty to disclose information which is material to patentability as defined in Title 37, Code of Federal Regulations, §1.56 which became available between the filing date of the prior application and the national or PCT international filing date of this application:

APPLICATION SERIAL NO.	FILING DATE	STATUS		
		PATENTED	PENDING	ABANDONED
PCT/US98/14715	July 21, 1998		X	

**POWER OF ATTORNEY:** As a named inventor, I hereby appoint S. Leslie Misrock (Reg. No. 18872), Harry C. Jones, III (Reg. No. 20280), Berj A. Terzian (Reg. No. 20060), Gerald J. Flintoft (Reg. No. 20823), David Weild, III (Reg. No. 21094), Jonathan A. Marshall (Reg. No. 24614), Barry D. Rein (Reg. No. 22411), Stanton T. Lawrence, III (Reg. No. 25736), Isaac Jarkovsky (Reg. No. 22713), Joseph V. Colaianni (Reg. No. 20019), Charles E. McKenney (Reg. No. 22795), Philip T. Shannon (Reg. No. 24278), Francis E. Morris (Reg. No. 24615), Charles E. Miller (Reg. No. 24576), Gidon D. Stern (Reg. No. 27469), John J. Lauter, Jr. (Reg. No. 27814), Brian M. Poissant (Reg. No. 28462), Brian D. Coggio (Reg. No. 27624), Rory J. Radding (Reg. No. 28749), Stephen J. Harbulak (Reg. No. 29166), Donald J. Goodell (Reg. No. 19766), James N. Palik (Reg. No. 25510), Thomas E. Friebel (Reg. No. 29258), Laura A. Coruzzi (Reg. No. 30742), Jennifer Gordon (Reg. No. 30753), Jon R. Stark (Reg. No. 30111), Allan A. Fanucci (Reg. No. 30256), Geraldine F. Baldwin (Reg. No. 31232), Victor N. Balancia (Reg. No. 31231), Samuel B. Abrams (Reg. No. 30605), Steven I. Wallach (Reg. No. 35402), Marcia H. Sundeen (Reg. No. 30893), Paul J. Zegger (Reg. No. 33821), Edmond R. Bannon (Reg. No. 32110), Bruce J. Barker (Reg. No. 33291), Adriane M. Antler (Reg. No. 32605), Thomas G. Rowan (Reg. No. 34419), James G. Markey (Reg. No. 31636), Thomas D. Kohler (Reg. No. 32797), Scott D. Stimpson (Reg. No. 33607), Ann L. Gisolfi (Reg. No. 31956), and Mark A. Farley (Reg. No. 33170), all of Pennie & Edmonds LLP, whose addresses are 1155 Avenue of the Americas, New York, New York 10036, 1667 K Street N.W., Washington, DC 20006 and 3300 Hillview Avenue, Palo Alto, CA 94304, and each of them, my attorneys, to prosecute this application, and to transact all business in the Patent and Trademark Office connected therewith.

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I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

SIGNATURE OF INVENTOR 201  Michal Eisenbach-Schwartz  DATE	SIGNATURE OF INVENTOR 202  Irun R. Cohen  DATE	SIGNATURE OF INVENTOR 203  Gila Moalem  DATE
SIGNATURE OF INVENTOR 204  DATE	SIGNATURE OF INVENTOR 205  DATE	SIGNATURE OF INVENTOR 206  DATE
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SIGNATURE OF INVENTOR 201  DATE	SIGNATURE OF INVENTOR 202  DATE	SIGNATURE OF INVENTOR 203  DATE